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(71) Applicant (for all designated States except US): THE SCRIPPS RESEARCH INSTITUTE [US/US]; 10666 North Torrey Pines Road, Mail Drop TPC-8, La Jolla, CA 92037 (US).

(72) Inventors; and

- (75) Inventors'Applicants (for US only): FENG, Lili [CN/US]; 7430 Mason Heights Lane, San Diego, CA 92126 (US). CHEN, Sizhong [CN/US]; 7896 Camino Kiosco, San Diego, CA 92122 (US). XIA, Yiyang [CN/US]; 7430 Mason Heights Lane, San Diego, CA 92126 (US).
- (74) Agents: OLSON, Ame, M.; Olson & Hierl, Ltd., 36th floor, 20 North Wacker Drive, Chicago, IL 60606 (US) et al.

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(54) Title: DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO REGULATING ENERGY MOBILIZATION WITH OB PROTEIN AND OB ANTIBODIES

(57) Abstract

Compositions comprising OB-R agonists and methods of treatment for conditions such as systemic inflammatory response syndrome are provided. One suitable OB-R agonist ligand is recombinant human OB protein, also known as leptin. Also provided are methods and compositions for the treatment of obesity and OB resistance. Assay methods and kits relating to these conditions are also included.

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# DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO REGULATING ENERGY MOBILIZATION WITH OB PROTEIN AND OB ANTIBODIES

# 5 Reference to Related Application

This application claims the benefit of U.S. Provisional Application S.N. 60/018,972, filed June 4, 1996, which is incorporated by reference, as are all references cited herein.

# Governmental Rights

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This invention was made with governmental support from the United States Government, National Institutes of Health, Grant DK20043; the United States Government has certain rights in the invention.

### Background of the Invention

The *obese* gene in human, rat and mouse encodes a protein hormone having an open reading frame 167 amino acid residues in length, called leptin, also known as OB protein or the *ob* gene product. Removal of the signal sequence yields a mature secreted 16 kilodalton protein that is 146 amino acid residues in length.

OB protein is produced primarily by adipocytes of white adipose tissue (WAT). OB protein is secreted directly into the extracellular space and travels through the blood stream. OB protein affects the cells of its target organs by binding to the OB receptor protein, OB-R, that is found on the extracellular surface of the plasma membrane of target cells. Binding of OB protein to OB-R activates the intracellular second messenger cascade of the JAK-STAT system, which is characteristic of activation of cytokine type I receptors.

OB protein is produced in adipocytes in proportion to the mass of stored fat, thereby providing a hormone signal for a lipostatic feedback circuit, which is mediated by the OB receptor. While OB proteins of different species show a close similarity in their sequences, the sequences of OB proteins are not closely similar to other types of proteins. For example, the human ob gene

sequence and its mouse homologue (85% sequence identity) have been reported to have no sequence similarity to other proteins of known structure (DiFrancesco, V., et al., Protein Topology Recognition from Secondary Structure Sequences: Application of the Hidden Markov Models to Alpha Class Proteins, <u>J. Mol. Biol.</u> **267:** 446-463 (1997) at page 457).

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Although OB protein is composed of a single peptide chain, an intrachain disulfide bond between cysteine 96 and cysteine 146 is required both to stabilize the conformation of the molecule and to confer in vivo biological activity (Rock, F.L., et al., The Leptin Haemopoietic Cytokine Fold is Stabilized by an Intrachain Disulfide Bond, Horm. Metab. Res. 28: 649-652 (1996)). It is believed that the special geometry of the A and D major helices must be maintained in order to dock to a conserved receptor trough in the receptor molecule, a requirement that produces structural similarity between OB proteins and cytokines in the face of negligible sequence conservation (Id. at 651.).

One accepted and successful animal model of human obesity is the genetically obese mouse bearing the recessive *obese* mutation (*ob/ob*). The mouse model reproduces not only the human obesity condition, but also develops non-insulin dependent diabetes mellitus (NIDDM, also known as type II diabetes mellitus). Homologous *obese* genes have been described in mouse, rat and human.

The mouse is also a widely accepted and successful model of sepsis, septic shock and systemic inflammatory response syndrome (SIRS), a term which describes the clinical syndrome of sepsis without regard to its cause. Simple models, involving a large bolus dose of lipopolysaccharide (LPS) administered to mice and using mortality as the primary outcome variable, are well suited for preliminary pharmacological studies of new drugs or other therapeutic agents (Fink, M.P. & Heard, S.O., Laboratory Models of Sepsis and Septic Shock, J. Surg. Res. 49: 186-196, 1990, at 188-189).

Both the *ob* gene (Zhang, Y., et a;., Positional cloning of the mouse *obese* gene and its human homologue, Nature 372: 425-432 (1994);

WO 97/46249 PCT/US97/09684 - 3 -

accession No. U18812, SEQ ID NO. 1) and its receptor (Tartaglia, L.A., et al., Identification and expression cloning of a Leptin Receptor, Cell 83: 1263-1271 (1995); Chen, H., et al., Evidence that the Diabetes Gene Encodes the Leptin Receptor: Identification of a Mutation in the Leptin Receptor Gene in db/db Mice Cell 84: 491-495 (1996); accession No. U46135, SEQ ID NO. 2) have been cloned. Shorter versions of the OB receptor, termed the OB-Ra, OB-Rc and OB-Re forms, are produced by alternative splicing of the OB-R mRNA (Lee, G. H., et al. Nature 379: 632-635 (1996)). The full length OB receptor is called the OB-Rb form.

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Neural activity in specific regions of the central nervous system (CNS), such as the hypothalamus, controls processes related to food intake and energy expenditure. The cloning of the OB protein gene and the OB receptor gene and the localization of OB receptor expression in the hypothalamus has provided supporting evidence for this view as well as suggesting possible mechanisms for relating food intake to stored fat reserves. The OB protein is produced by adipocytes in proportion to the mass of stored fat and, hence, it acts as the signal to a lipostat control circuit. This lipostat signal is transduced at the target cells by the OB receptor, OB-R, in the CNS, resulting changes in neural

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Metabolic derangement is an important characteristic of the host response to critical illness called the acute phase response that characterizes conditions such as sepsis and septic shock (Kushner, I. Ann. N.Y. Acad. Sci. 389: 39-48 (1982)). Hypothermia is a metabolic response that may be pertinent clinical prognostic factor in systemic inflammatory response syndrome in humans (Brivet, F., et al. Crit. Care Med. 22: 533-534 (1994)).

activity that regulate both food intake and metabolic rate.

There is a need for at least one disease marker for systemic inflammatory response syndrome (SIRS) and related conditions. Hereinafter, the term SIRS is used to denote sepsis, septic shock, sepsis syndrome, and related conditions. Disease markers have numerous functions. In this case, a marker for SIRS would be useful for predicting the development of SIRS, identifying patients with SIRS, predicting outcome, aiding timing and targeting of therapeutic interventions, and determining the pathogenesis of SIRS in patients (Parsons, P.E. & Moss, M. Early Detection and Markers of Sepsis, Clinics in Chest Medicine 17:199-212 (1996)).

## 5 Summary of the Invention

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Embodiments of the present invention include compositions and methods for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired, comprising administering a composition having a physiologically effective amount of at least one OB-R agonist ligand. Suitable OB-R agonist ligands include recombinant OB protein, peptide conformational analogs of human OB protein comprising conservative substitutions of amino acid residues and OB-related peptides. A preferred OB-R agonist ligand is recombinant human OB protein.

In another therapeutic embodiment, the beneficial aspects of OB-R agonist ligand administration are facilitated by a coordinated increase in the number of OB receptors (OB-R) produced by the administration of agents that OB-R expression inducers. Suitable OB-R expression inducers include lipopolysaccharide (LPS) and cytokines. Preferred cytokines are interleukin- $1\alpha$  (IL- $1\alpha$ ), interleukin- $1\beta$  (IL- $1\beta$ ), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin 6 (IL-6). Particularly preferred cytokines are IL-6 and IL- $1\beta$ .

In a further therapeutic embodiment, antibodies to OB protein are used as agents capable of blocking the effects of OB receptor activation, especially fat mobilization and increased energy utilization. Suitable antibodies to OB proteins may be polyclonal or monoclonal. Suitable antibodies comprise immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, i.e., molecules that contain an antibody combining site or paratope. Exemplary antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and those portions of an immunoglobulin molecule that contain the paratope, including those portions known in the art as Fab, Fab' and F(ab')<sub>2</sub>. This embodiment is suitable for

WO 97/46249 - 5 - PCT/US97/09684

treating metabolic derangement due to conditions such as anorexia, amenorrhea, cachexia and the like.

Antibodies to OB protein are also useful as an assay kit and method for detecting the level of OB protein in a patient. The level of OB protein in a patient is a disease marker that is useful for predicting the development of a condition, identifying patients with the condition, predicting outcome of the condition, aiding timing and targeting of therapeutic interventions, and determining the pathogenesis of the condition in patients. Conditions in which the level of OB protein is a useful marker are SIRS and related conditions such as sepsis and septic shock, as well as anorexia, amenorrhea, cachexia and the like.

## Brief Summary of the Drawings

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In the drawings:

Figure 1 is a representation of an autoradiogram showing the results of a ribonuclease (RNase) protection assay showing expression of total OB receptor (OB- $R_{(t)}$ ) in lung, kidney and liver at 0, 4, 8 or 24 hours, or 2 ( $D_2$ ), 3 ( $D_3$ ) or 5 ( $D_5$ ) days after intravenous injection of 5  $\mu$ g per gram of body weight as well as a dose-response study showing the relative effects on the liver the of injection of 0.05, 0.5 or 5  $\mu$ g of LPS per gram of body weight;

Figure 2 is a representation of an autoradiogram showing of the results of a RNase protection assay showing expression of OB-Rb in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and *ob/ob* mice (lane 4), compared to OB-R<sub>(m)</sub>) which represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

Figure 3 is a representation of an autoradiogram showing the results of a RNase protection assay showing expression of OB-Rc in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and ob/ob mice (lane 4), compared to OB-R<sub>(m)</sub>) which

represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

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Figure 4 is a representation of an autoradiogram showing the results of a RNase protection assay showing expression of OB-Re in normal liver (lane 1), and the LPS-treated liver at 24 hours (lane 2), and in the hypothalamus of normal control (lane 3) and *ob/ob* mice (lane 4), compared to OB-R<sub>(m)</sub>) which represents the mixture of OB-R forms partially protected by the designated nucleotide probes;

Figure 5 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB-R expression in mouse liver 24 hours after IL-6 (2.5  $\mu$ g per mouse), TNF- $\alpha$  (10  $\mu$ g per mouse) and IL-1 $\beta$  (5  $\mu$ g per mouse) injection;

Figure 6 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB-R mRNA levels in brain cortex, hypothalamus, and brain stem at various times after LPS injection (5  $\mu$ g per gram of body weight);

Figure 7 is a representation of an autoradiogram showing the results of a RNase protection assay showing OB mRNA expression in the adrenal gland (Adr) and white adipose tissue (WAT) at various times after LPS injection (5  $\mu$ g per gram of body weight);

Figure 8 is a graphical representation of the construction scheme for the vector pETM1 from a commercially available vector;

Figure 9 is a graphical representation of the weight gain induced by anti-OB antiserum injection in C57BL/6 mice which were given daily injections of anti-OB antiserum (antiOB) or preimmune rabbit sera (control), and whose body weight was measured 12 hours later, where data are expressed as mean  $\pm$  standard error of the mean (S.E.M), N=8;

Figure 10 is a graphical representation of the time course of the survival of mice that had received a LPS injection (6  $\mu$ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB,

WO 97/46249 PCT/US97/09684

N=16), preimmune rabbit serum (control, N=16), or one of three other unrelated rabbit antisera (anti-X, N=9: 3 treated with each antiserum);

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Figure 11 is a graphical representation of the time course of the survival of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N-16) after a LPS injection (10  $\mu$ g per gram of body weight);

Figure 12 is a graphical representation of the time course of the change in body temperature of mice that had received a LPS injection (6  $\mu$ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB, N=16, except at \* where N<16 due to mortality) or preimmune rabbit serum (control, N=16), data expressed as mean  $\pm$  S.E.M.;

Figure 13 is a graphical representation of the time course of the change in body temperature of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N-16, except at \* where N<16 due to mortality) after a LPS injection (10  $\mu$ g per gram of body weight), data expressed as mean  $\pm$  S.E.M.;

Figure 14 is a graphical representation of the time course of the change in body weight (percent of initial body weight) of mice that had received a LPS injection (6  $\mu$ g per gram of body weight) after pretreatment with either anti-OB protein antiserum (anti-OB, N=16) or preimmune rabbit serum (control, N=16, except at \* where N<16 due to mortality), data expressed as mean  $\pm$  S.E.M.;

Figure 15 is a graphical representation of the time course of the change in body weight (percent of initial body weight) of mice that were treated with OB protein (mOB, N=16) or vehicle (control, N-16, except at \* where N<16 due to mortality) after a LPS injection (10  $\mu$ g per gram of body weight), data expressed as mean  $\pm$  S.E.M.; and

Figure 16 is a representation of an autoradiogram showing the results of a RNase protection assay showing the expression of iNOS, IL-1 $\alpha$ , IL-1 $\beta$ , and TNF- $\alpha$  mRNAs in mouse.

WO 97/46249 - 8 - PCT/US97/09684

# Detailed Description of the Preferred Embodiments

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It has been found that substances that initiate or mediate SIRS, for example, LPS and several cytokines, induce the increased expression of OB-R in liver and other peripheral tissues. Thus, occupancy and activation of OB-R by an agonist ligand such as recombinant OB protein, OB-related peptides or peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues serves as a protective homeostatic mechanism in systemic inflammatory response syndrome conditions such as endotoxic shock, sepsis and septic shock. A preferred OB-R agonist ligand is recombinant OB protein. Suitable therapeutic human doses of recombinant OB protein are from about 1 micrograms per kilogram body weight. One preferable therapeutic human dose is about 10 micrograms per kilogram body weight.

While the regulation of energy homeostasis is essentially a function of the CNS, food intake and the majority of the energy expenditure take place in peripheral organs such as the liver. It has been found that the OB protein and the OB receptor have a functional involvement in peripheral energy homeostasis. In general, critical illness and trauma can dramatically alter metabolism, with the expression of the OB receptor changing in response to pathological stress. The expression of OB-R in liver and other peripheral organs, but not in the central nervous system, has now been shown to be induced by endotoxic shock produced intravenous injection of cytokines, such as, IL-1 $\beta$ , TNF- $\alpha$  and IL-6, as well as cytokine inducing agents such as LPS, into mice, an accepted animal model of SIRS and related conditions. OB protein, antibodies to OB protein, and OB-R expression inducers are useful for the diagnosis and treatment of conditions such as sepsis, systemic inflammatory response syndrome, cachexia and anorexia.

The administration of recombinant mouse OB protein to mice following OB-R induction with a normally lethal dose of LPS conferred complete resistance to LPS, resulting in survival. The OB-treated mice maintained a higher body temperature and displayed dramatic weight loss in contrast to control

counterparts. <u>In vivo</u> administration of OB antisera, on the other hand, elicited the opposite effects by blocking OB-mediated processes, thereby stimulating post-prandial food intake leading to rapid weight gain. Co-administration of LPS with a second <u>in vivo</u> treatment with OB antisera, however, resulted in 100% mortality as compared to animals treated with control antisera.

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OB protein, in mediating host responses to LPS-induced endotoxemia, exerts its protective effect primarily by initiating energy mobilization and heat production in critical conditions, the effect of which is proportional to the level of OB protein in the blood. By altering the levels of OB protein, the amount of energy mobilized to resist challenges induced by inflammatory agents is correspondingly altered, thereby effecting the ultimate inflammatory response.

Therefore, in view of the newly discovered physiological properties of OB protein and OB antibodies in regulating energy mobilization and consumption, the present invention describes both diagnostic and therapeutic methods relating to the use of recombinant human OB protein and antibodies thereto in modulating the effects of OB-R activation <u>in vivo</u>.

Diagnostic Applications

OB antibodies are useful for detecting the amount of OB present in sample taken from a patient. One preferred diagnostic embodiment is the use of OB antibodies for detecting the amount of OB present in a blood sample taken from a patient exhibiting a SIRS condition such as sepsis, septic shock, and the like. Such measurements of OB levels in vitro in blood sample by antibody detection is also indicated in various wasting conditions or syndromes associated with several disease states or syndromes, including anorexia, amenorrhea, cachexia, chronic inflammatory conditions, AIDS and AIDS-related conditions, as well as sepsis, septic shock, SIRS and the like. Alternatively, the determination of OB levels is useful in systemic inflammatory response syndromes (SIRS) that are characterized by an acute increase in inflammatory mediators, such as IL-1 $\beta$ , Il-6, TNF, LPS and the like. Such conditions are noted in preoperative patients

subject to fasting, in patients with acute injuries such as burns or trauma, in patients with SIRS, or with ongoing bacterial infections or those receiving TNF- $\alpha$  for treatment of tumors and in persons suffering from hypothermia.

While there is little sequence similarity between OB proteins and other molecules, the three-dimensional conformation of the OB protein molecule is analogous to that of several long-chain helical cytokines: four major alpha helix regions, A-D, connected by short loops and minor helical regions (Zhang, F., et. al., Crystal Structure of the *obese* protein leptin-E100, Nature 387: 206-209 (1997)).

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As used herein, a conformational analog of OB protein is a molecule having substantially the same conformational characteristics of its three-dimensional structure that are required for activation of the OB receptor. Examples of such conformational characteristics include the conformation of the A major helix, the conformation of the D major helix, and the disulfide bond that maintains the geometrical relationship between the A and D major helices. Thus, amino acid substitutions that conserve the conformational characteristics of the molecule, for example, in the loop regions connecting the major helices, would produce conformational analogs to OB protein.

Peptides derived from the region of the OB protein from amino acid residues 106 to 140, as short as about 15 amino acids long, have been shown to be effective in mimicking the action of full-length recombinant OB protein (Grasso, P., et al., In vivo Effects of Leptin-Related Synthetic Peptides on Body Weight and Food Intake in Female *ob/ob* Mice: Localization of Leptin Activity to Domains Between Amino Acid Residues 106 - 140, Endocrinology 138: 1413-1418 (1997)). As used herein, "OB-related peptides" refers to natural or synthetic peptides derived from the region of the OB protein from about amino acid residue 106 to about amino acid residue 140 and includes conservative amino acid residue substitutions.

The term "antibody" in its various grammatical forms is used herein to refer to immunoglobulin molecules and immunologically active portions

of immunoglobulin molecules, i.e., molecules that contain an antibody combining site or paratope. Exemplary antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and those portions of an immunoglobulin molecule that contain the paratope, including those portions known in the art as Fab, Fab' and  $F(ab')_2$ . An antibody composition of the present invention is characterized as containing antibody molecules that immunoreact with OB protein or portions thereof.

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An antibody composition of the present invention is typically produced by immunizing a mammal with a inoculum of OB protein or some fragment of OB protein, alone or in combination with a suitable adjuvant such as Freund's adjuvant, and thereby induce in the mammal antibody molecules having the appropriate immunospecificity. The antibody molecules are then collected from the mammal and isolated to the extent desired by well known techniques such as, for example, immunoaffinity chromatography. The antibody composition so produced can be used, inter alia, in the diagnostic methods and systems of the present invention or in the preparation of therapeutic compositions of the present invention.

Monoclonal antibody compositions can also be used with the present invention. A monoclonal antibody composition contains, within detectable limits, only one species of antibody combining site capable of effectively binding to OB protein. Thus, a monoclonal antibody composition of the present invention typically displays a single binding affinity for OB protein even though it may contain antibodies capable of binding proteins other than OB protein. Preferred monoclonal antibodies are those that bind to portions of the OB protein that are required for activation of the OB receptor, such as the A helix, the D helix, or regions of OB protein that maintain the relative positions of the A and D helices that are required for the activation of the OB receptor. Monoclonal antibodies against human OB protein have been described and their preparation was discussed in (Tsuruo, Y. et. al., Horm. Metab. Res. 28: 753-755 (1996).

Monoclonal antibodies are also supplied commercially from vendors on a custom

order basis (e.g., Alpha Diagnostic International, Inc., San Antonio, TX). Purified polyclonal anti-OB antibodies are commercially available from several sources (R&D Systems, Minneapolis, MN; Research Diagnostics, Inc., Flanders, N.J.; Linco Research, Inc., St. Charles, MO; Affinity BioReagents, Inc., Golden, CO).

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Determination of OB levels with OB antibodies is performed by assay methods, including ELISA, radioimmunoassay (RIA), Western blot analysis, and the like, that are familiar to one of ordinary skill in the art. The determined OB protein levels are then compared to normal levels for the state of the patient, e.g., fasting, time of day, body mass index (BMI), aerobic conditioning, gender, etc. For example, the normal range found for lean males at 8 a.m. was  $12.0 \pm 4.4$  ng/ml (Sinha, M.K., et al., Nocturnal Rise of Leptin in Lean, Obese, and Non-Insulin-dependent Diabetes Mellitus Subjects, J. Clin. Invest. 97: 1344-1347 (1996)). See, also Horn, R. et al., Radioimmunoassay for the detection of leptin in human serum, Exp. Clin. Endocrinol. Diabetes 104: 454-458 (1996); McGregor, G.P., et al., Radioimmunological Measurement of Leptin in Plasma of Obese and Diabetic Human Subjects, Endocrinology 137: 1501-1504 (1996). It has recently been found that OB protein is present in the circulation in both bound and free form, and that the ratio of the two forms is different in lean and obese subjects (Sinha, M.K., et al., Evidence of Free and Bound Leptin in Human Circulation. Studies in Lean and Obese Subjects and During Short-Term Fasting, J. Clin. Invest. 98: 1277-1282 (1996)). The relation of free and bound forms to OB protein biological activity can be considered in the context of OB protein assays.

In an alternative embodiment, immunohistochemical assay of OB-R receptor numbers are performed on tissue biopsy materials using standard protocols. A preferred tissue biopsy is liver biopsy.

# Therapeutic Applications

Embodiments of the present invention, including methods of administering to a patient compositions comprising OB-R agonist ligands are useful in treating conditions in which it is desirable to regulate or modify energy metabolism during a systemic inflammatory response. Suitable OB-R agonist ligands include recombinant OB protein, peptide conformational analogs of human OB protein comprising conservative substitutions of amino acid residues and OB-related peptides. A preferred OB-R agonist ligand is recombinant human OB protein. A suitable dosage range for recombinant human OB protein is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight. OB-related peptides are used in a dosage range from about 0.1 microgram per kilogram body weight to about 5 micrograms per kilogram body weight, adjusting the dosage to account for art-recognized differences in potency and solubility (Grasso, P. et al., (1997)).

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In a related embodiment, compositions comprising at least one OB-R expression inducer are useful for treating obesity and conditions in which there is an insufficient number of OB receptors, in which low copy number of OB receptors is a limiting factor or in which there is "OB resistance," i.e., a reduced effect of associated with a particular plasma concentration of OB protein.

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In general, the OB-R expression inducer is administered in an amount from about 0.003 to about 20 micrograms per kilogram body weight. Suitable OB-R expression inducers include therapeutic cytokines used in cancer therapy, such as IL-1 $\alpha$ , IL-1 $\beta$ , IL-6 and TNF- $\alpha$ . Suitable dosages and modes of administration are known in the art. For example, a suitable dosage ranges for IL-1 $\alpha$  is about 0.1 to about 6 micrograms/m²/day. A suitable dosage range for IL-1 $\beta$  is about 3 to about 200 nanograms/kg/day. A suitable dosage range for IL-6 is about 0.5 to about 20 micrograms/kg/day, with a preferred dosage range for IL-6 being about 1 to about 5 micrograms/kg/day. The therapeutic cytokines may be administered singly or in combination. The mode of administration may be intravenous infusion over an extended time period or a single intravenous or

subcutaneous injection. The daily dose may be administered as a single dose or divided into multiple dose given at intervals during the day.

In a further embodiment, to counteract the possible toxic side effects of OB-R expression inducers such as therapeutic cytokines, such substances are administered in a composition in combination with OB-R agonist ligands. Administration of such compositions is useful for conditions in with cytokines are normally administered for a therapeutic purpose such as tumor treatment, in order to provide effective protection by the OB-R agonist ligands from undesirable metabolic side effects. The up-regulation of the OB-R allows for the complete therapeutic effect mediated by OB-R agonist ligands such as OB protein. A suitable dosage range for recombinant human OB protein is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

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In an alternative embodiment, the present invention describes the administration of OB antisera, polyclonal or monoclonal, for treating conditions marked by increased OB and/or increased OB-R activity. Such conditions are various wasting conditions or syndromes associated with several disease states or syndromes, including anorexia, amenorrhea, cachexia, chronic inflammatory conditions, AIDS and AIDS-related conditions. A suitable dosage range for anti-OB protein antibodies is about 0.02 to about 15 milligrams / kg / day.

It has been found that OB protein is an important host defense factor against endotoxin stress. The protective effect of OB protein against endotoxin was not caused by suppressing the expression of major inflammatory mediators, since the mRNA levels of IL- $1\alpha$ , IL- $1\beta$ , TNF- $\alpha$ , and iNOS in lung and spleen were similar in all LPS-treated mice regardless of the experimental manipulations (Figure 16). A comparison of the four groups of mice revealed striking correlations among the OB protein available, the survival from endotoxin shock (Figures 10 and 11), the maintenance of body temperature (Figures 12 and 13), and the loss of body weight (Figures 14 and 15).

Anti-OB Ab-treated mice showed the least loss of body weight and had the most profound hypothermia even with a relatively low dose of LPS. Conversely, OB-treated mice receiving a high dose of LPS maintained a higher body temperature than those control mice that received less LPS. The OB-treated mice also had a greater weight loss than any reported in the literature (16% in the first 24 hours, compared with an average of 10% reported by other groups). The mice in these tests were age, sex and weight matched, fed the same diet, and, therefore, should have had very similar energy store. The different responses to endotoxemia described were likely due to differences in metabolic energy mobilization and dissipation, which, in turn, were attributed to the experimental manipulation of circulating OB protein levels.

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When the level of circulating OB protein is varied, the energy mobilized to resist endotoxin challenge is correspondingly altered, and the outcome of the host response to endotoxin stress is affected. ob/ob mice, lacking OB protein due to a mutation in the ob gene, were very sensitive to LPS insult: a dose as low as 2  $\mu$ g per gram of body weight caused a rapid fall of body temperature and death.

The results also suggest the existence of two pathways of thermogenesis and thermostasis. Anti-OB Ab treatment per se did not cause hypothermia in normal mice, suggesting that the thermostasis under non-pathological conditions was largely OB protein-independent. However, when given LPS, the anti-OB Ab-treated mice developed profound hypothermia, indicating that the thermogenesis in response to endotoxemia had become OB protein dependent. A corollary of this model is that genetic defects affecting the OB/OB-R pathway will have a severe hypothermic response to endotoxin. Indeed, *db/db* mice which carry a mutation in the OB-R gene, responded to a low dose of LPS injection in a manner very similar to that seen in *ob/ob* mice (data not shown), despite their increased level of OB expression.

# Example 1: Induction of OB-R Expression by Administration of LPS

The injection of LPS and cytokines, substances that are associated with sepsis, septic shock or SIRS caused increases in the expression of the OB receptor in peripheral organs such as liver, but not in brain.

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Intravenous injection of lipopolysaccharide (LPS), IL-1β, TNF-α and IL-6 to mice induced OB receptor expression in the liver and other peripheral organs, but not in the central nervous system (CNS). To investigate the functional significance of the increased OB-R expression, an anti-OB antiserum was used to neutralize endogenous OB protein in mice prior to an LPS injection. The neutralization of OB protein led to profound hypothermia, insignificant loss of body weight, and death in mice in response to an otherwise nonlethal dose of LPS. Conversely, mice administered recombinant mouse OB protein became more resistant to LPS and survived an otherwise lethal dose. The OB protein-treated mice maintained a relatively high body temperature and displayed a dramatic weight loss. These results suggest that OB protein may promote energy mobilization to compensate for the increased energy consumption in endotoxemia, and that the OB/OB-R pathway may play an important role in critical host responses to inflammatory stress.

Methods:

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In general, standard techniques or published modifications were used; see, generally, Sambrook, J., et al., Molecular Cloning A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory Press (1989). C57BL/6 mice, 5-8 weeks of age and 17-20 g of weight, were injected intravenously with either LPS (5  $\mu$ g per gram of body weight, List Biological Laboratory, Campbell, CA), IL-1 $\beta$  (R&D Systems, Minneapolis, MN), IL-6 (2.5  $\mu$ g, Pharmingen, San Diego, CA), or TNF- $\alpha$  (a gift from Genentech, San Francisco, CA). Animals were sacrificed at 0, 4, 8 or 24 hours, or 2, 3 or 5 days after the injection.

Tissue from various organs, including brain, liver and kidney, was dissected and snap-frozen in liquid nitrogen. Total RNA was prepared from the frozen tissues by a single-step method (Chomczynski, P. & Sacchi, N., Single-

Step Method of RNA Isolation by Acid Guanidinium Thiocyanate-Phenol-Chloroform Extraction, <u>Analytical Biochemistry</u> **162**: 156-159 (1987).

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The RNase protection assays were carried out as previously described (Feng, L., et al., Alternative Splicing of the NC1 Domain of the Human  $\alpha 3$ (IV) Collagen Gene <u>J. Biol. Chem.</u> **269**: 2342-2348 (1994); Xia, Y., et al., LPS-Induced MCP-1, IL-1 $\beta$ , and TNF- $\alpha$  mRNA Expression in Isolated Erythrocyte-Perfused Rat Kidney, Am. J. Physiol. 264: F774-F780 (1993)). A... ten microgram aliquot of total RNA pooled from three similarly treated mice was used for each sample in the RNase protection assay. The stored pooled samples were dissolved in 10  $\mu$ l of 80% formamide, 0.4 M NaCl, 1 mM EDTA, and 40 mM piperazine-N,N'-bis(2-ethanesulfonic acid), heated to 85 degrees Celsius for 5 minutes. Each ten microgram sample was then hybridized with about 1 x 10<sup>5</sup> cpm (counts per minute) of the appropriate [32P]UTP-labeled antisense riboprobe at 55 degrees Celsius for at least 10 hours. The unhybridized RNA was then digested with 50 unit/ml RNase T1 (GIBCO/BRL, Gaithersburg, MD) and 24 μg/ml RNase A at 30 degrees Celsius for one hour. The RNase was then digested with 625  $\mu$ g/ml proteinase K (Boehringer Mannheim, Indianapolis, IN) for 30 minutes at 37 degrees Celsius. After phenol-chloroform extraction and sodium-acetate-ethanol precipitation, the protected hybridized RNA was denatured and electrophoresed on a 10% polyacrylamide gel. The gels were transferred to 3M Whatman filter paper, dried and exposed to Kodak X-Omat film. The resulting autoradiograms were developed in a Kodak X-Omat processor were used only for qualitative screening.

Radioactivity due to hybridization of target sequences with <sup>32</sup>P-labeled riboprobes was quantified by scanning the gels on an AMBIS radioanalytic scanning system (AMBIS Systems, San Diego, CA).

An OB-Rb cDNA probe (from base 2548 to base 2835 of OB-Rb, Gen-Bank™ Accession No. U46135) was subcloned from a full-length mouse OB-Rb cDNA. The full-length mouse cDNA for the OB-R long form (OB-Rb)

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was cloned from a mouse hypothalamus cDNA library (Stratagene, La Jolla, CA), and the sequence was verified against that of U46135.

The full-length mouse cDNA for the OB-R short form (OB-Ra) was cloned from a mouse lung cDNA Library (Stratagene, La Jolla, CA). A 224 bp probe that included base 1250 to base 1474 (as indicated on the OB-Rb sequence) of OB-Ra was used for the RNase protection assay. This fragment, which comprises a sequence that is shared by all variants of OB-R, was used as a probe for the total level of OB-R (OBR<sub>(t)</sub>).

The expression of other forms of OB-R mRNA was analyzed using selective probes for the respective different forms of OB-R. The designated probes provided full protection to their corresponding OB-R forms and partial protection for other OB-R forms. A probe derived from L32 (33-126, Gen-Bank™ Accession No. XO6483), a housekeeping gene encoding ribosomal protein, was used as a control.

OB-Rc and OB-Re probes were cloned by reverse-transcription-PCR (RT-PCR) of total liver RNA from LPS-treated C57BL/6 mice. Protocols for RT-PCR are known in the art (for example, pages 15-13 - 15-15 of Ausubel, F.M., et al., Short Protocols in Molecular Biology, 2nd Edition, John Wiley and Sons, New York, (1992)).

One suitable protocol for RT-PCR is a modification of that previously described (Feng, L., et al., <u>J. Biol. Chem.</u> **269:** 2342-2348, 1994). The primers used in RT-PCR are listed in Table 1, below. Primer oligonucleotides were synthesized using an ABI model 380B synthesizer (Applied BioSystems, Foster City, Ca).

Table 1: PCR Primers

		Sequence	
	OB-Rc sense	5'-GCTATCGACAAGCAGCAGAAT-3'	(SEQ ID NO. 8)
5	OB-Rc antisense	5'-TGAACACAACATAAAGCCC-3'	(SEQ ID NO. 9)
	OB-Re sense	5'-TGTTATATCTGGTTATTATTGAATGG-3'	(SEQ ID NO. 10)
10	OB-Re antisense	5'-CATTAAATGATTTATTATCAGAATTGC-3'	(SEQ ID NO. 11)

First strand cDNA synthesis was performed using total liver RNA from LPS-treated C57BL/6 mice and murine leukemia virus reverse transcriptase with a random hexanucleotide primer. The 100  $\mu$ l reaction mixture contained standard enzyme buffer, 5  $\mu$ g of total RNA, 20 units of RNasin (RNase inhibitor), 500 pmol of hexanucleotide primer, 10 mM dithiothreitol, 1 mM of each dNTP, with 200 units of reverse transcriptase. Each reaction mixture was heated to 95 degrees Celsius for 10 minutes. PCR was then performed with separate aliquots of the reaction mixture with the appropriate primers for 35 cycles, using 60 degrees Celsius for annealing.

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The cDNA segments used to generate riboprobes were excised by the appropriate restriction endonucleases and subcloned into the multiple cloning site of a standard transcription vector. Suitable transcription vectors include a vector chosen from the pGEM series (Promega, Madison, WI). Labelled single stranded riboprobes were synthesized using standard in vitro transcription protocols, either those provided by the manufacturer or other standard protocols (e.g. Ausubel, F.M., et al., pages 4-18 - 4-21) with the appropriate (e.g., SP6 or T7) bacteriophage RNA polymerase. The riboprobes contained regions corresponding to the vector polylinker in addition to the region corresponding to the target sequence, and thus were longer than the protected bands. The mouse ribosomal L32 gene, a constitutively expressed "housekeeping" gene, was used throughout the study as a control.

WO 97/46249 - 20 -

### Results:

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When LPS was administered to C57BL/6 mice, a strong induction of total OB-R expression (OB-R<sub>(0)</sub>) was detected in a number of peripheral organs (Figure 1), but not in several areas of the central nervous system, such as the hypothalamus, that are known to express OB-R (compare Figure 1 and Figure 6). The increased expression of OB-R was most prominent in the liver, the major site of metabolic regulation. The increase of OB-R mRNA expression in the liver was LPS dose-dependent, and peaked between 24 and 48 hours post-LPS injection (Figure 1).

Unexpectedly, RNase protection assays using probes specific for alternatively spliced forms of OB-R mRNA revealed that the long form, OB-Rb, was also induced in the liver to a level comparable to that found in the *ob/ob* mouse hypothalamus and greater than that of the lean control mouse hypothalamus (Figure 2). However, the majority of hepatic OB-R were the OB-Ra, OB-Rc (Figure 3) and OB-Re (Figure 4) forms. OB-Rd expression in the liver was undetectable (data not shown).

In addition to LPS, OB-R expression was induced by the cytokines IL-6, IL-1 $\alpha$  and TNF- $\alpha$  (Figure 5). Contrary to a recent report (12), we found no detectable increase in OB expression in white adipose tissue in LPS-treated mice, but detected a distinct induction of OB mRNA expression in the adrenal gland (Figure 4). No OB mRNA expression was found in the brain, heart, lung, liver, kidney, spleen, muscle, stomach, duodenum, jejunum, ileum, or colon of LPS-treated mice (data not shown).

### Example 2: Production of Recombinant OB Protein

Recombinant OB protein was expressed in <u>E. coli</u> using a prokaryotic expression vector and extracted from inclusion bodies. Other vectors and host cells systems, including eukaryotic cells, are known in the art and also suitable for the expression of OB protein. See, generally, Ausubel, F.M., et al. <u>Short Protocols in Molecular Biology</u>, 2nd Ed., pages 16-1 to 16-89.

The coding region of mouse OB cDNA (65-619, Gen-Bank™ Accession No.U18812) was cloned by RT-PCR of total RNA from C57BL/6 white adipose tissue. The coding region was subcloned in expression vector, pETM1 (Feng, L., et al., J. Biol. Chem. 269: 2342-2348, 1994), to express a His-tagged recombinant mouse OB protein. The construction of pETM1 from the commercially available vector pET-11a (Novagen, Madison, WI) is illustrated on Figure 8.

After the expression of OB protein is induced, the bacteria were harvested and the inclusion bodies were extracted with a buffer containing 6M urea. The extract was loaded on a Ni-NTA affinity column (Qiagen, Chatsworth, CA) and the purification procedure was carried out as previously described (Feng, et al.(1994)). The protein was refolded on the column by adding refolding buffer containing 5 mM CaCl<sub>2</sub>/20 mM Tris/0.2 NaCl with an urea gradient of 4 M - 0.5 M at a rate of about 0.5 ml/minute. After refolding, the protein was eluted with 80 mM imidazole/5 mM CaCl<sub>2</sub>/20 mM Tris/0.2 NaCl/ 0.5 mM urea and then dialyzed against phosphate-buffered saline (PBS). Polyclonal antibodies was raised by immunizing a rabbit with the recombinant mouse OB and Freund's adjuvant using standard procedures. Antiserum was used in the following examples.

### 20 Example 3: Effects of Anti-OB Antibodies on Metabolism

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Intravenous administration of antibodies directed against OB protein effectively opposes the effects of endogenous OB protein.

Recombinant mouse OB protein was produced in an <u>E. coli</u> expression system as described in Example 2, and was used to generate rabbit polyclonal anti-OB antibody. The antibody, when injected intravenously into mice, stimulated food intake, leading to rapid weight gain, and thus was effective in blocking OB protein function. The results are shown in Figure 9.

Female C57BL/6 mice, 6-8 weeks of age and 15-17 g of weight, were group housed four per cage and adapted to a 12:12 hour light: dark cycle (light from 6:00 to 18:00). Mice were given a daily intravenous injection of 0.2

ml anti-OB antiserum or preimmune rabbit sera ("vehicle") at 10 p.m., after their initial dark phase food intake. Their body weight was measured at 10 a.m. the next day.

The weight gain induced by anti-OB antiserum is illustrated in Figure 9. Data are expressed as mean  $\pm$  S.E.M. (N=8). While the weight of the control group remained essentially constant over the week, the anti-OB treated group showed a weight increase at the first weighing, which continued for the entire study period.

# Example 4: Effects of Anti-OB Antibodies and Recombinant OB-Protein on Response to Endotoxic Shock

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The fact that the OB-R variants induced in the liver were predominantly short forms raised the question of functional relevance of the hepatic OB-R expression. OB-Rb is the main form expressed in the hypothalamus, while the choroid plexus expresses only OB-Ra. That the mutation in *db/db* mice affects OB-Rb, but not OB-Ra, suggests that OB-Rb is crucial for regulating food intake and OB-Ra may act as an OB protein transporter. Accordingly, the prominent expression of OB-R in the liver could initiate intracellular signal transduction or, alternatively, mediate the clearance of OB protein. We found that administration of neutralizing anti-OB antibody (Ab) or OB protein to LPS-treated mice distinguished between the two alternatives.

Male C57BL/6 mice, 5-8 weeks of age and 17-21 g of body weight, were used for this study. For antisera treatment, mice were given an i.v. injection of 200  $\mu$ l rabbit antisera. LPS at a dose of 6  $\mu$ g per gram of body weight was then co-injected with a second dose of anti-OB antibody to the pretreated mice. LPS was dissolved in antisera at a concentration of 0.6 mg/ml and was injected intravenously 4 hours after the initial antisera treatment. Food was retrieved from mouse cages during the 4-hour pretreatment period to prevent any food intake differences resulting from anti-OB antiserum-induced hyperphagia, and was added back after LPS injection.

While this dose of LPS was not lethal to C57BL/6 mice treated with preimmune rabbit serum, all the mice in the anti-OB Ab-treated group died within 40 hours. Figure 10 illustrates the results from three groups of mice: those pretreated with anti-OB antiserum (anti-OB, N=16), preimmune rabbit sera control, N=16), or three other unrelated rabbit antisera ("Anti-X", N=3 for each antiserum). This LPS sensitizing effect was specific for anti-OB Ab, since mice treated similarly with three other unrelated antibodies ("Anti-X") all survived (Figure 10).

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In comparison, mice treated with OB protein (5  $\mu$ g per gram of body weight mOB, N=16) were able to survive a higher dose of LPS (10  $\mu$ g per gram of body weight) that was fatal to the control group of mice receiving the vehicle alone (control, N=16). Mouse OB protein and LPS were prepared in saline at a concentration of 0.5 mg/ml and 1 mg/ml, respectively, and injected intravenously into the mice. A vehicle solution used for OB protein dialysis was injected into control mice. To eliminate any circadian effect, experiments at different days were all started at the same hours. Mice were examined at 4-hour intervals post LPS injection for the first 24 hours, and survival was monitored for 7 days following the LPS injection. The results are shown in Figure 11. The dose of LPS killed all the mice in the control group within 24 hours. However, in the experimental group, OB protein treatment conferred mice complete resistance to this dose of endotoxin. The OB-treated mice displayed noticeably less severe symptoms of endotoxemia, remaining alert and responsive to touch and other manipulation, and recovering quickly.

The effects of both OB protein and anti-OB antibodies on body temperature and body weight were monitored in the same groups of mice. Core body temperature measurements were made by insertion of a thermistor probe (Yellow Springs Instrument, Yellow Springs, Ohio) into the colon, 1.5 cm beyond the rectum. Body weight measurements were made on a portable digital balance (Ohaus, Florham Park, NJ).

The mice receiving anti-OB antibodies, which ultimately died, showed lower body temperature (Figure 12) and less weight loss (Figure 14) than the corresponding control group than survived. Conversely, the mice receiving OB-protein, and which survived the endotoxic shock, showed higher body temperature (Figure 13) and more weight loss (Figure 15) than the corresponding control group which succumbed.

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While administration of both OB protein and anti-OB antibodies had significant effects on survival, body weight and body temperature, there was little effect on the expression of iNOS, IL- $1\alpha$ , IL- $1\beta$ , and TNF- $\alpha$  mRNAs in lung and spleen (Figure 16). Mice were treated as described in above (Figures 10-15).

RNase protection assays were performed as described in Example 1. Each sample was 5  $\mu$ g total RNA was used for each sample in the RNase protection assay. Riboprobes were produced as described in Example 1 based on the following cDNA fragments: IL-l $\alpha$  (from base 172 to base 366, Accession No. X01450, SEQ ID NO. 3), IL-1 $\beta$  (from base 500 to base 671, Accession No. M15131, SEQ ID NO. 4), TNF- $\alpha$  (from base 428 to base 557, Accession No. M11731, SEQ ID NO. 5), mouse iNOS (from base 2404 to base 2698, Accession No. M92649, SEQ ID NO. 6), and L32 (from base 33 to base 126, Accession No. X064383, SEQ ID NO. 7).

Relatively little change in the pattern of expression of these markers was observed (Figure 16), suggesting that these the protective actions of OB proteins are direct and not indirect and mediated by these cytokines.

The foregoing is intended to be illustrative of the present invention, but not limiting. Numerous variations and modifications of the present invention may be effected without departing from the true spirit and scope of the invention.

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Feng, Lili Chen, Sizhong Xia, Yiyang
  - (ii) TITLE OF INVENTION: DIAGNOSTIC AND THERAPEUTIC METHODS RELATED TO REGULATING ENERGY MOBILIZATION WITH OB PROTEIN AND OB ANTIBODIES
  - (iii) NUMBER OF SEQUENCES: 11
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: Olson & Hierl, Ltd.
    - (B) STREET: 20 North Wacker Drive, 36th Floor
    - (C) CITY: Chicago
    - (D) STATE: IL
    - (E) COUNTRY: US
    - (F) ZIP: 60606
    - (v) COMPUTER READABLE FORM:
      - (A) MEDIUM TYPE: Floppy disk
      - (B) COMPUTER: IBM PC compatible
      - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
      - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
  - (vi) CURRENT APPLICATION DATA:
    - (A) APPLICATION NUMBER:
    - (B) FILING DATE: 04-JUN-1997
    - (C) CLASSIFICATION:
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 60/018,972
    - (B) FILING DATE: 04-JUN-1996
  - (viii) ATTORNEY/AGENT INFORMATION:
    - (A) NAME: Olson, Arne M
    - (B) REGISTRATION NUMBER: 30,203
    - (C) REFERENCE/DOCKET NUMBER: TSRI540.1PCT
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: 312-580-1180
      - (B) TELEFAX: 312-580-1189
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2793 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO

# (vi) ORIGINAL SOURCE: (A) ORGANISM: Mus musculus

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

60	AGGAAAATGT	GATCCCAGGG	AGAAGAAGAA	GCAAGGTGCA	TCCAGCAGCT	GGATCCCTGC
120	GTTCAAGCAG	TCTGTCTTAT	TTTGGTCCTA	TTCCTGTGGC	CCTGTGTCGG	GCTGGAGACC
180	GTCACCAGGA	CAAGACCATT	AAACCCTCAT	GATGACACCA	GAAAGTCCAG	TGCCTATCCA
240	GGCTTGGACT	GAGGGTCACT	CCGCCAAGCA	CAGTCGGTAT	TTCACACACG	TCAATGACAT
300	CTGGCAGTCT	GGACCAGACT	TGTCCAAGAT	ATTCTGAGTT	GCTTCACCCC	TCATTCCTGG
360	AATGACCTGG	GCAGATAGCC	AAAATGTGCT	CTGCCTTCCC	CCTCACCAGC	ATCAACAGGT
420	CTGCCTCAGA	GAGCTGCTCC	CCTTCTCCAA	CATCTGCTGG	AGACCTCCTC	AGAATCTCCG
480	CTCTACTCCA	GGAAGCCTCA	ATGGCGTCCT	GAGAGCCTGG	GCAGAAGCCA	CCAGTGGCCT
540	CAACAGTTGG	GGACATTCTT	GCTCTCTGCA	AGGCTGCAGG	GGCTTTGAGC	CAGAGGTGGT
600	CATGTAGAGG	TCCCAAGAAT	GCCACCAGGC	AGTTTCAAAG	TGAATGCTGA	ATGTTAGCCC
660	ACATCCATCA	CCATGTGCAC	GAGAAGAGAG	GGGTCTTCAG	TGGCTTCCAG	GAAGAAACCT
720	ATGCTTGACT	TGACTCCACA	TCCAAAGGCA	AGACCACCCA	TCCCTCCTGT	TTCATTTCTC
780	GGGACTCTCA	GCCTGCAGAG	GGAGGGGCCA	ATGAGCACAA	ACACAACTTC	CAAGTTATCC
840	CCACCTGCTC	CCTCCATGTC	CATCCCATCC	AGATAAGAGC	CAGCAAGTAG	CCTAGTTCTT
900	TGAGGTAGGG	CCAGGAGAGG	TCGCTGCGGC	GGTACACGCT	TTCCTCCGTG	CGGGTACATG
960	CTGCATCCAC	ACCGTGAAGG	CTTTGGGAGC	GTCTCAGAGT	CCTTTGGGCT	ATGGGTAGAG
1020	ATTCTGCATT	TTATTTATTT	ATGGAAGCAC	GCAGCACACG	AAACTCCCAA	ACACAGCTGG
1080	GTCAGCCAGG	GGCTTTGGGG	AGCTTTTTCA	GCAAGGCATC	TGGATCTGAA	CTATTTTGGA
1140	CGAGGCAAAC	TGGGTCTGCC	ATCCTATTGA	GCTGCTTTCA	CTCCTGGGGT	ATGAGGAAGG
1200	TATGCAGGTA	AACAAGAGTC	GGATCTTCCA	AGGAAGGTTG	AGTGACTGGA	CTAATTTTTG
1260	TCTATCCAAA	TGTGACTGAC	TTGTTTCTAT	GTGACTGGTT	TTGACCTCTG	GCGCTCAAGA
1320	CAGATGAATT	TTATCAAAAG	GGCTAGGTTA	CGGGAGCATA	GCGGCATTGC	CACGTTTGCA
1380	AGAGGGAGGG	AGGATGTGTT	CTGAGGGTAG	CTATGTGCAC	TAATATGTAT	TTGTCAAGTG
1440	TGAAAGGGTG	TAGGCTTTTC	ATATGTGTGG	TCTGAATTAC	GGAAGTGTTC	TGAAGGATCC
1500	AAAGGAGTTG	TGAAAAGGAC	TGTGGCTTTG	GGCCACATAG	TTACCTCTGT	AGGCATTTTC
1560	GCTACAGGCC	AGGGGCTAAA	GCACCCTTGG	AGTGTACCAG	GAACATTTGG	ACTCTTTCCG
1620	CAGTGAGCCC	ACATTTGAGA	TGAGGGCCCC	GCTCAGGGAG	ATATTGCTGA	TTTTGTTGGC
1680	CACAATGCGT	GGGTTGATCT	AGGTTGTCCA	TAGATCTCCA	GTCCCTGGTG	CAAGAAAAGG

TTCTTAAGCA	GGTAGACGTT	TGCATGCCAA	TATGTGGTTC	TCATCTGATT	GGTTCATCCA	1740
AAGTAGAACC	CTGTCTCCCA	CCCATTCTGT	GGGGAGTTTT	GTTCCAGTGG	GAATGAGAAA	1800
TCACTTAGCA	GATGGTCCTG	AGCCCTGGGC	CAGCACTGCT	GAGGAAGTGC	CAGGGCCCCA	1860
GGCCAGGCTG	CCAGAATTGC	CCTTCGGGCT	GGAGGATGAA	CAAAGGGGCT	TGGGTTTTTC	1920
CATCACCCCT	GCACCCTATG	TCACCATCAA	ACTGGGGGGC	AGATCAGTGA	GAGGACACTT	1980
GATGGAAAGC	AATACACTTT	AAGACTGAGC	ACAGTTTCGT	GCTCAGCTCT	GTCTGGTGCT	2040
GTGAGCTAGA	GAAGCTCACC	ACATACATAT	AAAAATCAGA	GGCTCATGTC	CCTGTGGTTA	2100
GACCCTACTC	GCGGCGGTGT	ACTCCACCAC	AGCAGCACCG	CACCGCTGGA	AGTACAGTGC	2160
TGTCTTCAAC	AGGTGTGAAA	GAACCTGAGC	TGAGGGTGAC	AGTGCCCAGG	GGAACCCTGC	2220
TTGCAGTCTA	TTGCATTTAC	ATACCGCATT	TCAGGGCACA	TTAGCATCCA	CTCCTATGGT	2280
AGCACACTGT	TGACAATAGG	ACAAGGGATA	GGGGTTGACT	ATCCCTTATC	CAAAATGCTT	2340
GGGACTAGAA	GAGTTTTGGA	TTTTAGAGTC	TTTTCAGGCA	TAGGTATATT	TGAGTATATA	2400
TAAAATGAGA	TATCTTGGGG	ATGGGGCCCA	AGTATAAACA	TGAAGTTCAT	TTATATTTCA	2460
TAATACCGTA	TAGACACTGC	TTGAAGTGTA	GTTTTATACA	GTGTTTTAAA	TAACGTTGTA	2520
TGCATGAAAG	ACGTTTTTAC	AGCATGAACC	TGTCTACTCA	TGCCAGCACT	CAAAAACCTT	2580
GGGGTTTTGG	AGCAGTTTGG	ATCTTGGGTT	TTCTGTTAAG	AGATGGTTAG	CTTATACCTA	2640
AAACCATAAT	GGCAAACAGG	CTGCAGGACC	AGACTGGATC	CTCAGCCCTG	AAGTGTGCCC	2700
TTCCAGCCAG	GTCATACCCT	GTGGAGGTGA	GCGGGATCAG	GTTTTGTGGT	GCTAAGAGAG	2760
GAGTTGGAGG	TAGATTTTGG	AGGATCTGAG	GGC			2793

### (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3862 base pairs

  - (B) TYPE: nucleic acid (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Mus musculus
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GTCGACCCAC GCGTCCGGAG GAATCGTTCT GCAAATCCAG GTGTACACCT CTGAAGAAAG 60 ATGATGTGTC AGAAATTCTA TGTGGTTTTG TTACACTGGG AATTTCTTTA TGTGATAGCT 120

GCACTTAACC	TGGCATATCC	AATCTCTCCC	TGGAAATTTA	AGTTGTTTTG	TGGACCACCG	180
AACACAACCG	ATGACTCCTT	TCTCTCACCT	GCTGGAGCCC	CAAACAATGC	CTCGGCTTTG	240
AAGGGGGCTT	CTGAAGCAAT	TGTTGAAGCT	TTAATTTAAA	CAAGTGGTAT	CTACGTTCCT	300
GAGTTATCCA	AAACAGTCTT	CCACTGTTGC	TTTGGGAATG	AGCAAGGTCA	AAACTGCTCT	360
GCACTCACAG	ACAACACTGA	AGGGAAGACA	CTGGCTTCAG	TAGTGAAGGC	TTCAGTTTTT	420
CGCCAGCTAG	GTGTAAACTG	GGACATAGAG	TGCTGGATGA	AAGGGGACTT	GACATTATTC	480
ATCTGTCATA	TGGAGCCATT	ACCTAAGAAC	CCCTTCAAGA	ATTATGACTC	TAAGGTCCAT	540
CTTTTATATG	ATCTGCCTGA	AGTCATAGAT	GATTCGCCTC	TGCCCCCACT	GAAAGACAGC	600
TTTCAGACTG	TCCAATGCAA	CTGCAGTCTT	CGGGGATGTG	AATGTCATGT	GCCGGTACCC	660
AGAGCCAAAC	TCAACTACGC	TCTTCTGATG	TATTTGGAAA	TCACATCTGC	CGGTGTGAGT	720
TTTCAGTCAC	CTCTGATGTC	ACTGCAGCCC	ATGCTTGTTG	TGAAACCCGA	TCCACCCTTA	780
GGTTTGCATA	TGGAAGTCAC	AGATGATGGT	AATTTAAAGA	TTTCTTGGGA	CAGCCAAACA	840
ATGGCACCAT	TTCCGCTTCA	ATATCAGGTG	AAATATTTAG	AGAATTCTAC	AATTGTAAGA	900
GAGGCTGCTG	AAATTGTCTC	AGCTACATCT	CTGCTGGTAG	ACAGTGTGCT	TCCTGGATCT	960
TCATATGAGG	TCCAGGTGAG	GAGCAAGAGA	CTGGATGGTT	CAGGAGTCTG	GAGTGACTGG	1020
AGTTCACCTC	AAGTCTTTAC	CACACAAGAT	GTTGTGTATT	TTCCACCCAA	AATTCTGACT	1080
AGTGTTGGAT	CGAATGCTTC	TTTTCATTGC	ATCTACAAAA	ACGAAAACCA	GATTATCTCC	1140
TCAAAACAGA	TAGTTTGGTG	GAGGAATCTA	GCTGAGAAAA	TCCCTGAGAT	ACAGTACAGC	1200
ATTGTGAGTG	ACCGAGTTAG	CAAAGTTACC	TTCTCCAACC	TGAAAGCCAC	CAGACCTCGA	1260
GGGAAGTTTA	CCTATGACGC	AGTGTAĊTGC	TGCAATGAGC	AGGCGTGCCA	TCACCGCTAT	1320
GCTGAATTAT	ACGTGATCGA	TGTCAATATC	AATATATCAT	GTGAAACTGA	CGGGTACTTA	1380
ACTAAAATGA	CTTGCAGATG	GTCACCCAGC	ACAATCCAAT	CACTAGTGGG	AAGCACTGTG	1440
CAGCTGAGGT	ATCACAGGCG	CAGCCTGTAT	TGTCCTGATA	GTCCATCTAT	TCATCCTACG	1500
TCTGAGCCCA	AAAACTGCGT	CTTACAGAGA	GACGGCTTTT	ATGAATGTGT	TTTCCAGCCA	1560
ATCTTTCTAT	TATCTGGCTA	TACAATGTGG	ATCAGGATCA	ACCATTCTTT	AGGTTCACTT	1620
GACTCGCCAC	CAACGTGTGT	CCTTCCTGAC	TCCGTAGTAA	AACCACTACC	TCCATCTAAC	1680
GTAAAAGCAG	AGATTACTGT	AAACACTGGA	TTATTGAAAG	TATCTTGGGA	AAAGCCAGTC	1740
TTTCCGGAGA	ATAACCTTCA	ATTCCAGATT	CGATATGGCT	TAAGTGGAAA	AGAAATACAA	1800
TGGAAGACAC	ATGAGGTATT	CGATGCAAAG	TCAAAGTCTG	CCAGCCTGCT	GGTGTCAGAC	1860
CTCTGTGCAG	TCTATGTGGT	CCAGGTTCGC	TGCCGGCGGT	TGGATGGACT	AGGATATTGG	1920
AGTAATTGGA	GCAGTCCAGC	CTATACGCTT	GTCATGGATG	TAAAAGTTCC	TATGAGAGGG	1980

CCTGAATTTT	GGAGAAAAT	GGATGGGGAC	GTTACTAAAA	AGGAGAGAAA	TGTCACCTTG	2040
CTTTGGAAGC	CCCTGACGAA	AAATGACTCA	CTGTGTAGTG	TGAGGAGGTA	CGTGGTGAAG	2100
CATCGTACTG	CCCACAATGG	GACGTGGTCA	GAAGATGTGG	GAAATCGGAC	CAATCTCACT	2160
TTCCTGTGGA	CAGAACCAGC	GCACACTGTT	ACAGTTCTGG	CTGTCAATTC	CCTCGGCGCT	2220
TCCCTTGTGA	ATTTTAACCT	TACCTTCTCA	TGGCCCATGA	GTAAAGTGAG	TGCTGTGGAG	2280
TCACTCAGTG	CTTATCCCCT	GAGCAGCAGC	TGTGTCATCC	TTTCCTGGAC	ACTGTCACCT	2340
GATGATTATA	GTCTGTTATA	TCTGGTTATT	GAATGGAAGA	TCCTTAATGA	AGATGATGGA	2400
ATGAAGTGGC	TTAGAATTCC	CTCGAATGTT	AAAAAGTTTT	ATATCCACGA	TAATTTTATT	2460
CCCATCGAGA	AATATCAGTT	TAGTCTTTAC	CCAGTATTTA	TGGAAGGAGT	TGGAAAACCA	2520
AAGATAATTA	ATGGTTTCAC	CAAAGATGCT	ATCGACAAGC	AGCAGAATGA	CGCAGGGCTG	2580
TATGTCATTG	TACCCATAAT	TATTTCCTCT	TGTGTCCTAC	TGCTCGGAAC	ACTGTTAATT	2640
TCACACCAGA	GAATGAAAAA	GTTGTTTTGG	GACGATGTTC	CAAACCCCAA	GAATTGTTCC	2700
TGGGCACAAG	GACTGAATTT	CCAAAAGCCT	GAAACATTTG	AGCATCTTTT	TACCAAGCAT	2760
GCAGAATCAG	TGATATTTGG	TCCTCTTCTT	CTGGAGCCTG	AACCCATTTC	AGAAGAAATC	2820
AGTGTCGATA	CAGCTTGGAA	AAATAAAGAT	GAGATGGTCC	CAGCAGCTAT	GGTCTCCCTT	2880
CTTTTGACCA	CACCAGACCC	TGAAAGCAGT	TCTATTTGTA	TTAGTGACCA	GTGTAACAGT	2940
GCTAACTTCT	CTGGGTCTCA	GAGCACCCAG	GTAACCTGTG	AGGATGAGTG	TCAGAGACAA	3000
CCCTCAGTTA	AATATGCAAC	TCTGGTCAGC	AACGATAAAC	TAGTGGAAAC	TGATGAAGAG	3060
CAAGGGTTTA	TCCATAGTCC	TGTCAGCAAC	TGCATCTCCA	GTAATCATTC	CCCACTGAGG	3120
CAGTCTTTCT	CTAGCAGCTC	CTGGGAGACA	GAGGCCCAGA	CATTTTTCCT	TTTATCAGAC	3180
CAGCAACCCA	CCATGATTTC	ACCACAACTT	TCATTCTCGG	GGTTGGATGA	GCTTTTGGAA	3240
CTGGAGGGAA	GTTTTCCTGA	AGAAAATCAC	AGGGAGAAGT	CTGTCTGTTA	TCTAGGAGTC	3300
ACCTCCGTCA	ACAGAAGAGA	GAGTGGTGTG	CTTTTGACTG	GTGAGGCAGG	AATCCTGTGC	3360
ACATTCCCAG	CCCAGTGTCT	GTTCAGTGAC	ATCAGGATCC	TCCAGGAGAG	ATGCTCACAC	3420
TTTGTAGAAA	ATAATTTGAG	TTTAGGGACC	TCTGGTGAGA	ACTTTGTACC	TTACATGCCC	3480
CAATTTCAAA	CCTGTTCCAC	GCACAGTCAC	AAGATAATGG	AGAATAAGAT	GTGTGACTTA	3540
ACTGTGTAAT	CTCATCCAAG	AAGCCTCAAG	GTTCCATTCC	AGTAGAGCCT	GTCATGTATA	3600
ATGTGTTCTT	TTATTGTTGT	GGATGTGGGA	GACAAGTGTC	AGAATCTAGT	GTGAAAATGA	3660
TTGTTTCCAA	ACTAAGTGTG	TCTATTTTCT	CTCAGTAATA	CAATGAAACA	TATGAGGAAG	3720
CCCTCATTAA	TCTAGTAATG	TAGATGGACT	CTTACTGAAT	ATATTCCCAA	GATACTTGGG	3780
GAAGTCTCCC	TAATTCTAGC	AAATAAAT	CCCAGGAATA	GAACTACTAA	ACACTGAATC	3840

### TGGAAAAAA AAAAAAAAA AG

3862

# (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1974 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Mus musculus

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AAGTCTCCAG	GGCAGAGAGG	GAGTCAACTC	ATTGGCGCTT	GAGTCGGCAA	AGAAATCAAG	60
ATGGCCAAAG	TTCCTGACTT	GTTTGAAGAC	CTAAAGAACT	GTTACAGTGA	AAACGAAGAC	120
TACAGTTCTG	CCATTGACCA	TCTCTCTCTG	AATCAGAAAT	CCTTCTATGA	TGCAAGCTAT	180
GGCTCACTTC	ATGAGACTTG	CACAGATCAG	TTTGTATCTC	TGAGAACCTC	TGAAACGTCA	240
AAGATGTCCA	ACTTCACCTT	CAAGGAGAGC	CGGGTGACAG	TATCAGCAAC	GTCAAGCAAC	300
GGGAAGATTC	TGAAGAAGAG	ACGGCTGAGT	TTCAGTGAGA	CCTTCACTGA	AGATGACCTG	360
CAGTCCATAA	CCCATGATCT	GGAAGAGACC	ATCCAACCCA	GATCAGCACC	TTACACCTAC	420
CAGAGTGATT	TGAGATACAA	ACTGATGAAG	CTCGTCAGGC	AGAAGTTTGT	CATGAATGAT	480
TCCCTCAACC	AAACTATATA	TCAGGATGTG	GACAAACACT	ATCTCAGCAC	CACTTGGTTA	540
AATGACCTGC	AACAGGAAGT	AAAATTTGAC	ATGTATGCCT	ACTCGTCGGG	AGGAGACGAC	600
TCTAAATATC	CTGTTACTCT	AAAAATCTCA	GATTCACAAC	TGTTCGTGAG	CGCTCAAGGA	660
GAAGACCAGC	CCGTGTTGCT	GAAGGAGTTG	CCAGAAACAC	CAAAACTCAT	CACAGGTAGT	720
GAGACCGACC	TCATTTTCTT	CTGGAAAAGT	ATCAACTCTA	AGAACTACTT	CACATCAGCT	780
GCTTATCCAG	AGCTGTTTAT	TGCCACCAAA	GAACAAAGTC	GGGTGCACCT	GGCACGGGGA	840
CTGCCCTCTA	TGACAGACTT	CCAGATATCA	TAAAAGCAGC	CTTATTTCGG	GAGTCTATTC	900
ACTTGGGAAG	TGCTGACAGT	CTGTATGTAC	CATGTACAGG	AACCTTCCTC	ACCCTGAGTC	960
ACTTGCACAG	CATGTGCTGA	GTCTCTGTAA	TTCTAAATGA	ATGTTTACCC	TCTTTGTAAG	1020
AGAAGAGCAA	ACCCTAGTGG	AGCCACCCCG	ACATATGATA	CTATCTGTTA	TTTTAAAGAG	1080
TACCCTATAG	TTTGCTCAGT	ACTAATCATT	TTAATTACTA	TTCTGCATGG	CATTCTTAGG	1140
AGGATCAAAA	AGACTCTACA	CATATTACAG	ATGGGTTAAC	AAAGGGATAA	AACAACTGAA	1200

AAGCACACTC	AATGCATTTG	GAATATAAAT	TCACAGACCA	ATCTCACTGT	GCACCTTCGG	1260
CTTCAAAATG	CCAGTTGAGT	AGGATAAAGG	TATAAGAACT	TAATGCTGTC	ATTTTCAAAA	1320
GGAAGGGGAC	AATAGCTACA	TCTTTCCTAC	CTCAGTGGGT	TTTACTCCAG	TGAGATCATT	1380
TGGATGAAAT	CCTCCTGTAA	CAGACCTCAA	GAAGGAGACA	GACTGTTGAA	TGTTATTTTT	1440
AAGTTATTTT	ATATATGTAT	TTATAAATAT	ATTTATGATA	ATTATATTAT	TTATGGAACA	1500
TCCTTAAATC	CTCTGAGCTT	GACAGGCATC	CTCACAGCAG	GATTTTCTAG	GTGGTCAGTT	1560
AGATATAGTT	TCCTCTAGAG	CACCATGCTA	CAGACTTTAC	ACTTTTTCCA	CAGCCACGAA	1620
GCTCTCTGTA	CATTCCTGTA	CTTGGGAGCC	CTTTCATCAT	GATCTTAATC	TGTACTGTTT	1680
ACTTTGTTCA	TCTAAAATGA	TAATTGAGTC	AGTCTTTTTC	CCTCCCATCC	TTAAAGCTGT	1740
CTGGGTATTC	TTACATCATT	CAGTCTCACC	TGTAACTAAC	ACCAACCATC	TAAAGATGGA	1800
AAGAGCTTAA	CTGTGACAAC	CACATCACTG	TTACCTGAAG	TTTCTTTTCT	AGAATGTAAT	1860
CAGTGTTTCC	CCTGGATTCC	AATTTTTTTT	TCAAACCACA	GTATCATGTA	ACTATCAACA	1920
ATAACAATCA	ACTCATTATT	ATTAATCATA	AAATAAATTA	ACAAGTTTGA	GCTG	1974

### (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1339 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Mus musculus

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

TGCAGGGTTC	GAGGCCTAAT	AGGCTCATCT	GGGATCCTCT	CCAGCCAAGC	TTCCTTGTGC	60
AAGTGTCTGA	AGCAGCTATG	GCAACTGTTC	CTGAACTCAA	CTGTGAAATG	CCACCTTTTG	120
ACAGTGATGA	GAATGACCTG	TTCTTTGAAG	TTGACGGACC	CCAAAAGATG	AAGGGCTGCT	180
TCCAAACCTT	TGACCTGGGC	TGTCCAGATG	AGAGCATCCA	GCTTCAAATC	TCACAGCAGC	240
ACATCAACAA	GAGCTTCAGG	CAGGCAGTAT	CACTCATTGT	GGCTGTGGAG	AAGCTGTGGC	300
AGCTACCTGT	GTCTTTCCCG	TGGACCTTCC	AGGATGAGGA	CATGAGCACC	TTCTTTTCCT	360
TCATCTTTGA	AGAAGAGCCC	ATCCTCTGTG	ACTCATGGGA	TGATGATGAT	AACCTGCTGG	420
TGTGTGACGT	TCCCATTAGA	CAGCTGCACT	ACAGGCTCCG	AGATGAACAA	CAAAAAAGCC	480

TCGTGCTGTC	GGACCCATAT	GAGCTGAAAG	CTCTCCACCT	CAATGGACAG	AATATCAACC	540
AACAAGTGAT	ATTCTCCATG	AGCTTTGTAC	AAGGAGAACC	AAGCAACGAC	AAAATACCTG	600
TGGCCTTGGG	CCTCAAAGGA	AAGAATCTAT	ACCTGTCCTG	TGTAATGAAA	GACGGCACAC	660
CCACCCTGCA	GCTGGAGAGT	GTGGATCCCA	AGCAATACCC	AAAGAAGAAG	ATGGAAAAGC	720
GGTTTGTCTT	CAACAAGATA	GAAGTCAAGA	GCAAAGTGGA	GTTTGAGTCT	GCAGAGTTCC	780
CCAACTGGTA	CATCAGCACC	TCACAAGCAG	AGCACAAGCC	TGTCTTCCTG	GGAAACAACA	840
GTGGTCAGGA	CATAATTGAC	TTCACCATGG	AATCTGTGTC	TTCCTAAAGT	ATGGGCTGGA	900
CTGTTTCTAA	TGCCTTCCCC	AGGGCATGTG	AAGGAGCTCC	CTTGTCATGA	ATGAGCAGAC	960
AGCTCAATCT	CTAGGACACT	CCTTAGTCCT	CGGCCAAGAC	AGGTCGCTCA	GGGTCACAAG	1020
AAACCATGGC	ACATTCTGTT	CAAAGAGAGC	CTGTGTTTCC	TCCTTGCCTC	TGATGGGCAA	1080
CCACTTACCT	ATTTATTTAT	GTATTTATTG	ATTGGTTGAT	CTATTTAAGT	TGATTCAAGG	1140
GGACATTAGG	CAGCACTCTC	TAGAACAGAA	CCTAGCTGTC	AACGTGTGGG	GGATGAATTG	1200
GTCATAGCCT	TGCACTTGAG	GTCTTTCATT	GAAGCTGAGA	ATAAATAGGT	TCCTATAATA	1260
TGGATGAGAA	TTTTTATGAA	TGAAGCATTA	GCACATTGCT	TTGATGAGTA	TGAAATAAAT	1320
TTCATTAAAC	AAACAAACA					1339

### (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1629 base pairs
    (B) TYPE: nucleic acid
    (C) STRANDEDNESS: double

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Mus musculus

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GCTGAGGGAC	TAGCCAGGAG	GGAGAACAGA	AACTCCAGAA	CATCCTGGAA	ATAGCTCCCA	60
GAAAAGCAAG	CAGCCAACCA	GGCAGGTTCT	GTCCCTTTCA	CTCACTGGCC	CAAGGCGCCA	120
CATCTCCCTC	CAGAAAAGAC	ACCATGAGCA	CAGAAAGCAT	GATCCGCGAC	GTGGAACTGG	180
CAGAAGAGGC	ACTCCCCAA	AAGATGGGGG	GCTTCCAGAA	CTCCAGGCGG	TGCCTATGTC	240
TCAGCCTCTT	CTCATTCCTG	CTTGTGGCAG	GGGCCACCAC	GCTCTTCTGT	CTACTGAACT	300
TCGGGGTGAT	CGGTCCCCAA	AGGGATGAGA	AGTTCCCAAA	TGGCCTCCCT	CTCATCAGTT	360

CTATGGCCCA	GACCCTCACA	CTCAGATCAT	CTTCTCAAAA	TTCGAGTGAC	AAGCCTGTAG	420
CCCACGTCGT	AGCAAACCAC	CAAGTGGAGG	AGCAGCTGGA	GTGGCTGAGC	CAGCGCGCCA	480
ACGCCCTCCT	GGCCAACGGC	ATGGATCTCA	AAGACAACCA	ACTAGTGGTG	CCAGCCGATG	540
GGTTGTACCT	TGTCTACTCC	CAGGTTCTCT	TCAAGGGACA	AGGCTGCCCC	GACTACGTGC	600
TCCTCACCCA	CACCGTCAGC	CGATTTGCTA	TCTCATACCA	GGAGAAAGTC	AACCTCCTCT	660
CTGCCGTCAA	GAGCCCCTGC	CCCAAGGACA	CCCCTGAGGG	GGCTGAGCTC	AAACCCTGGT	720
ATGAGCCCAT	ATACCTGGGA	GGAGTCTTCC	AGCTGGAGAA	GGGGGACCAA	CTCAGCGCTG	780
AGGTCAATCT	GCCCAAGTAC	TTAGACTTTG	CGGAGTCCGG	GCAGGTCTAC	TTTGGAGTCA	840
TTGCTCTGTG	AAGGGAATGG	GTGTTCATCC	ATTCTCTACC	CAGCCCCCAC	TCTGACCCCT	900
TTACTCTGAC	CCCTTTATTG	TCTACTCCTC	AGAGCCCCCA	GTCTGTGTCC	TTCTAACTTA	960
GAAAGGGGAT	TATGGCTCAG	AGTCCAACTC	TGTGCTCAGA	GCTTTCAACA	ACTACTCAGA	1020
AACACAAGAT	GCTGGGACAG	TGACCTGGAC	TGTGGGCCTC	TCATGCACCA	CCACCCACGG	1080
AATCGAGAAA	GAGCTATCAA	TCTGGAATTC	ACTGGAGCCT	CGAATGTCCA	TTCCTGAGTT	1140
CTGCAAAGGG	AGAGTGGTCA	GGTTGCCTCT	GTCTCAGAAT	GAGGCTGGAT	AAGATCTCAG	1200
GCCTTCCTAC	CTTCAGACCT	TTCCAGACTC	TTCCCTGAGG	TGCAATGCAC	AGCCTTCCTC	1260
ACAGAGCCAG	CCCCCTCTA	TTTATATTTG	CACTTATTAT	TTATTATTŢA	TTTATTATTT	1320
ATTTATTTGC	TTATGAATGT	ATTTATTTGG	AAGGCCGGGG	TGTCCTGGAG	GACCCAGTGT	1380
GGGAAGCTGT	CTTCAGACAG	ACATGTTTTC	TGTGAAAACG	GAGCTGAGCT	GTCCCCACCT	1440
GGCCTCTCTA	CCTTGTTGCC	TCCTCTTTTG	CTTATGTTTA	ааасаааата	TTTATCTAAC	1500
CCAATTGTCT	TAATAACGCT	GATTTGGTGA	CCAGGCTGTC	GCTACATCAC	TGAACCTCTG	1560
CTCCCCACGG	GAGCCGTGAC	TGTAATTGCC	CTACGGGTCA	TTGAGAGAAA	TAAAGATCGC	1620
ттссалалас						1629

# (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 4110 base pairs
    (B) TYPE: nucleic acid
    (C) STRANDEDNESS: double
    (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Mus musculus

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

					CACCTTGGTG	60
					TGTTCACTCC	120
ACGGAGTAGC	CTAGTCAACT	GCAAGAGAAC	GGAGAACGTT	GGATTTGGAG	CAGAAGTGCA	180
AAGTCTCAGA	CATGGCTTGC	CCCTGGAAGT	TTCTCTTCAA	AGTCAAATCC	TACCAAAGTG	240
ACCTGAAAGA	GGAAAAGGAC	ATTAACAACA	ACGTGAAGAA	AACCCCTTGT	GCTGTTCTCA	300
GCCCAACAAT	ACAAGATGAC	CCTAAGAGTC	ACCAAAATGG	CTCCCCGCAG	CTCCTCACTG	360
GGACAGCACA	GAATGTTCCA	GAATCCCTGG	ACAAGCTGCA	TGTGACATCG	ACCCGTCCAC	420
AGTATGTGAG	GATCAAAAAC	TGGGGCAGTG	GAGAGATTTT	GCATGACACT	CTTCACCACA	480
AGGCCACATC	GGATTTCACT	TGCAAGTCCA	AGTCTTGCTT	GGGGTCCATC	ATGAACCCCA	540
AGAGTTTGAC	CAGAGGACCC	AGAGACAAGC	CTACCCCTCT	GGAGGAGCTC	CTGCCTCATG	600
CCATTGAGTT	CATCAACCAG	TATTATGGCT	CCTTTAAAGA	GGCAAAAATA	GAGGAACATC	660
TGGCCAGGCT	GGAAGCTGTA	ACAAAGGAAA	TAGAAACAAC	AGGAACCTAC	CAGCTCACTC	720
TGGATGAGCT	CATCTTTGCC	ACCAAGATGG	CCTGGAGGAA	TGTCCCTCGC	TGCATCGGCA	780
GGATCCAGTG	GTCCAACCTG	CAGGTCTTTG	ACGCTCGGAA	CTGTAGCACA	GCACAGGAAA	840
TGTTTCAGCA	CATCTGCAGA	CACATACTTT	ATGCCACCAA	CAATGGCAAC	ATCAGGTCGG	900
CCATCACTGT	GTTCCCCCAG	CGGAGTGACG	GCAAACATGA	CTTCAGGCTC	TGGAATTCAC	960
AGCTCATCCG	GTACGCTGGC	TACCAGATGC	CCGATGGCAC	CATCAGAGGG	GATGCTGCCA	1020
CCTTGGAGTT	CACCCAGTTG	TGCATCGACC	TAGGCTGGAA	GCCCCGCTAT	GGCCGCTTTG	1080
ATGTGCTGCC	TCTGGTCTTG	CAAGCTGATG	GTCAAGATCC	AGAGGTCTTT	GAAATCCCTC	1140
CTGATCTTGT	GTTGGAGGTG	ACCATGGAGC	ATCCCAAGTA	CGAGTGGTTC	CAGGAGCTCG	1200
GGTTGAAGTG	GTATGCACTG	CCTGCCGTGG	CCAACATGCT	ACTGGAGGTG	GGTGGCCTCG	1260
AATTCCCAGC	CTGCCCCTTC	AATGGTTGGT	ACATGGGCAC	CGAGATTGGA	GTTCGAGACT	1320
TCTGTGACAC	ACAGCGCTAC	AACATCCTGG	AGGAAGTGGG	CCGAAGGATG	GGCCTGGAGA	1380
CCCACACACT	GGCCTCCCTC	TGGAAAGACC	GGGCTGTCAC	GGAGATCAAT	GTGGCTGTGC	1440
TCCATAGTTT	CCAGAAGCAG	AATGTGACCA	TCATGGACCA	CCACACAGCC	TCAGAGTCCT	1500
TCATGAAGCA	CATGCAGAAT	GAGTACCGGG	CCCGTGGAGG	CTGCCCGGCA	GACTGGATTT	1560
GGCTGGTCCC	TCCAGTGTCT	GGGAGCATCA	CCCCTGTGTT	CCACCAGGAG	ATGTTGAACT	1620
ATGTCCTATC	TCCATTCTAC	TACTACCAGA	TCGAGCCCTG	GAAGACCCAC	ATCTGGCAGA	1680
ATGAGAAGCT	GAGGCCCAGG	AGGAGAGAGA	TCCGATTTAG	AGTCTTGGTG	AAAGTGGTGT	1740
TCTTTGCTTC	CATGCTAATG	CGAAAGGTCA	TGGCTTCACG	GGTCAGAGCC	ACAGTCCTCT	1800

TTGCTACTGA	GACAGGGAAG	TCTGAAGCAC	TAGCCAGGGA	CCTGGCCACC	TTGTTCAGCT	1860
ACGCCTTCAA	CACCAAGGTT	GTCTGCATGG	ACCAGTATAA	GGCAAGCACC	TTGGAAGAGG	1920
AGCAACTACT	GCTGGTGGTG	ACAAGCACAT	TTGGGAATGG	AGACTGTCCC	AGCAATGGGC	1980
AGACTCTGAA	GAAATCTCTG	TTCATGCTTA	GAGAACTCAA	CCACACCTTC	AGGTATGCTG	2040
TGTTTGGCCT	TGGCTCCAGC	ATGTACCCTC	AGTTCTGCGC	CTTTGCTCAT	GACATCGACC	2100
AGAAGCTGTC	CCACCTGGGA	GCCTCTCAGC	TTGCCCCAAC	AGGAGAAGGG	GACGAACTCA	2160
GTGGGCAGGA	GGATGCCTTC	CGCAGCTGGG	CTGTACAAAC	CTTCCGGGCA	GCCTGTGAGA	2220
CCTTTGATGT	CCGAAGCAAA	CATCACATTC	AGATCCCGAA	ACGCTTCACT	TCCAATGCAA	2280
CATGGGAGCC	ACAGCAATAT	AGGCTCATCC	AGAGCCCGGA	GCCTTTAGAC	CTCAACAGAG	2340
CCCTCAGCAG	CATCCATGCA	AAGAACGTGT	TTACCATGAG	GCTGAAATCC	CAGCAGAATC	2400
TGCAGAGTGA	AAAGTCCAGC	CGCACCACCC	TCCTCGTTCA	GCTCACCTTC	GAGGGCAGCC	2460
GAGGGCCCAG	CTACCTGCCT	GGGGAACACC	TGGGGATCTT	CCCAGGCAAC	CAGACCGCCC	2520
TGGTGCAGGG	AATCTTGGAG	CGAGTTGTGG	ATTGTCCTAC	ACCACACCAA	ACTGTGTGCC	2580
TGGAGGTTCT	GGATGAGAGC	GGCAGCTACT	GGGTCAAAGA	CAAGAGGCTG	CCCCCTGCT	2640
CACTCAGCCA	AGCCCTCACC	TACTTCCTGG	ACATTACGAC	CCCTCCCACC	CAGCTGCAGC	2700
TCCACAAGCT	GGCTCGCTTT	GGCACGGACG	AGACGGATAG	GCAGAGATTG	GAGGCCTTGT	2760
GTCAGCCCTC	AGAGTACAAT	GACTGGAAGT	TCAGCAACAA	CCCCACGTTC	CTGGAGGTGC	2820
TTGAAGAGTT	CCCTTCCTTG	CATGTGCCCG	CTGCCTTCCT	GCTGTCGCAG	CTCCCTATCT	2880
TGAAGCCCCG	CTACTACTCC	ATCAGCTCCT	CCCAGGACCA	CACCCCTCG	GAGGTTCACC	2940
TCACTGTGGC	CGTGGTCACC	TACCGCACCC	GAGATGGTCA	GGGTCCCCTG	CACCATGGTG	3000
TCTGCAGCAC	TTGGATCAGG	AACCTGAAGC	CCCAGGACCC	AGTGCCCTGC	TTTGTGCGAA	3060
GTGTCAGTGG	CTTCCAGCTC	CCTGAGGACC	CCTCCCAGCC	TTGCATCCTC	ATTGGGCCTG	3120
GTACGGGCAT	TGCTCCCTTC	CGAAGTTTCT	GGCAGCAGCG	GCTCCATGAC	TCCCAGCACA	3180
AAGGGCTCAA	AGGAGGCCGC	ATGAGCTTGG	TGTTTGGGTG	CCGGCACCCG	GAGGAGGACC	3240
ACCTCTATCA	GGAAGAAATG	CAGGAGATGG	TCCGCAAGAG	AGTGCTGTTC	CAGGTGCACA	3300
CAGGCTACTC	CCGGCTGCCC	GGCAAACCCA	AGGTCTACGT	TCAGGACATC	CTGCAAAAGC	3360
AGCTGGCCAA	TGAGGTACTC	AGCGTTCTCC	ACGGGGAGCA	GGGCCACCTC	TACATTTGCG	3420
GAGATGTGCG	CATGGCTCGG	GATGTGGCTA	CCACATTGAA	GAAGCTGGTG	GCCACCAAGC	3480
TGAACTTGAG	CGAGGAGCAG	GTGGAAGACT	ATTTCTTCCA	GCTCAAGAGC	CAGAAACGTT	3540
ATCATGAAGA	TATCTTCGGT	GCAGTCTTTT	CCTATGGGGC	AAAAAAGGGC	AGCGCCTTGG	3600
AGGAGCCCAA	AGCCACGAGG	CTCTGACAGC	CCAGAGTTCC	AGCTTCTGGC	ACTGAGTAAA	3660

GATAATGGTG	AGGGGCTTGG	GGAGACAGCG	AAATGCAATC	CCCCCCAAGC	CCCTCATGTC	3720
ATTCCCCCCT	CCTCCACCCT	ACCAAGTAGT	ATTGTATTAT	TGTGGACTAC	TAAATCTCTC	3780
TCCTCTCCTC	CCTCCCCTCT	CTCCCTTTCC	TCCCTTCTTC	TCCACTCCCC	AGCTCCCTCC	3840
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ACCTCCTGAC	TGAAGCACTT	TGGGTGACCA	CCAGGAGGCA	CCATGCCGCC	GCTCTAATAC	3960
TTAGCTGCAC	TATGTACAGA	TATTTATACT	TCATATTTAA	GAAAACAGAT	ACTTTTGTCT	4020
ACTCCCAATG	ATGGCTTGGG	CCTTTCCTGT	ATAATTCCTT	GATGAAAAAT	ATTTATATAA	4080
AATACATTTT	ATTTTAATCA	АААААААА				4110

#### (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 465 base pairs (B) TYPE: nucleic acid

  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Rattus norvegicus
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

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AAGTTCATCA	GGCACCAGTC	GGACCGATAT	GTGAAAATTA	AGCGAAACTG	GCGGAAACCC	120
AGAGGCATCG	ACAACAGGGT	GCGGAGAAGA	TTCAAGGGCC	AGATCCTGAT	GCCCAACATT	180
GGTTACGGGA	GTAACAAGAA	AACCAAGCAC	ATGCTGCCTA	GCGGCTTCCG	GAAGTTTCTG	240
GTCCACAATG	TCAAGGAGCT	GGAAGTGCTG	CTGATGTGCA	ACAAATCTTA	CTGTGCTGAG	300
ATTGCTCACA	ATGTGTCCTC	TAAGAACCGA	AAAGCCATCG	TAGAAAGAGC	AGCACAGCTG	360
GCCATCAGAG	TCACCAATCC	CAACGCCAGG	CTACGCAGCG	AAGAGAATGA	ATAGATGGCT	420
TGTGTGCCTG	TTTTGTGTTC	AAATAAAACC	ACAAAAACTG	CCAAA		465

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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 21 base pairs
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    (C) STRANDEDNESS: single

  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA

. (iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (A) ORGANISM: Mus musculus	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GCTATCGA	CA AGCAGCAGAA T	21
(2) INFO	RMATION FOR SEQ ID NO:9:	
(i)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 22 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
(ii)	MOLECULE TYPE: cDNA	
(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (A) ORGANISM: Mus musculus	
	SEQUENCE DESCRIPTION: SEQ ID NO:9:	
		22
	RMATION FOR SEQ ID NO:10:	
(1)	SEQUENCE CHARACTERISTICS:  (A) LENGTH: 26 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
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(iii)	HYPOTHETICAL: NO	
(iv)	ANTI-SENSE: NO	
(vi)	ORIGINAL SOURCE: (A) ORGANISM: Mus musculus	`
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TGTTATATC	T GGTTATTATT GAATGG	26
(2) INFOR	MATION FOR SEQ ID NO:11:	
(i)	SEQUENCE CHARACTERISTICS:	

- (A) LENGTH: 27 base pairs
- (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Mus musculus
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CATTAAATGA TTTATTATCA GAATTGC

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#### WE CLAIM:

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- 1. A method for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired, comprising administering a composition having a physiologically effective amount of at least one OB-R agonist ligand.
- 2. The method of claim 1 wherein the OB-R agonist ligand is.. recombinant human OB protein.
- 3. The method of claim 2 wherein the amount of recombinant human OB protein administered is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.
  - 4. The method of claim 1 wherein the OB-R agonist ligand is a peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues.
- 5. The method of claim 1 wherein the OB-R agonist ligand is an OB-related peptide.
  - 6. The method of claim 1 wherein the condition is sepsis.
  - 7. The method of claim 1 wherein the condition is systemic inflammatory response syndrome.
- 8. A composition for treating a patient having a condition in which regulating energy metabolism during a systemic inflammatory response is desired, comprising a physiologically effective amount of at least one OB-R agonist ligand.
- 9. The composition of claim 8 wherein the OB-R agonist ligand is recombinant human OB protein.
  - 10. The composition of claim 9 wherein the amount of recombinant human OB protein per dose is from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

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- 11. The composition of claim 8 wherein the OB-R agonist ligand is a peptide conformational analog of human OB protein comprising conservative substitutions of amino acid residues.
- 12. The composition of claim 8 wherein the OB-R agonist ligand is an OB-related peptide.
  - 13. The composition of claim 8 wherein the condition is sepsis.
  - 14. The composition of claim 8 wherein the condition is systemic inflammatory response syndrome.
  - 15. A composition for the amelioration of the toxicity of therapeutic cytokines comprising a physiologically effective amount of an OB-R agonist ligand.
    - 16. The composition of claim 15 wherein the OB-R agonist ligand is recombinant human OB protein.
    - 17. The composition of claim 15 wherein the amount of recombinant human OB protein per dose is 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.
    - 18. A method for the treatment of a patient having obesity comprising the steps of:

administering at least one OB-R expression inducer; and administering a physiologically effective amount of an OB-R agonist ligand.

- 19. The method of claim 18 wherein the OB-R expression inducer is a compound chosen from the group consisting of LPS, IL-1 $\alpha$ , IL-1 $\beta$ , TNF- $\alpha$  and IL-6.
- 20. The method of claim 18 wherein the OB-R expression inducer and the OB-R agonist ligand are administered at a different times.
- 21. The method of claim 18 wherein the OB-R expression inducer is administered in an amount from about 0.003 to about 20 micrograms per kilogram body weight.

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- 22. The method of claim 18 wherein the OB-R agonist ligand is administered in an amount from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.
- The method of claim 18 wherein the OB-R agonist ligand is recombinant human OB protein.
  - 24. The method of claim 23 wherein the recombinant human OB protein is administered in an amount from about 1 micrograms per kilogram. body weight to about 50 micrograms per kilogram body weight.
  - 25. The method of claim 18 wherein the OB-R expression inducer is IL-6.
    - 26. The method of claim 25 wherein IL-6 is administered in an amount from about 0.5 to about 20 micrograms per kilogram body weight.
    - 27. A method for the treatment of a patient having a condition characterized by OB resistance, comprising the steps of:
  - administering IL-6 in an amount from about 0.5 to about 20 micrograms per kilogram body weight; and

administering recombinant human OB protein in an amount from about 1 microgram per kilogram body weight to about 50 micrograms per kilogram body weight.

- 28. A composition suitable for the treatment of obesity comprising:
  - at least one therapeutic cytokine capable of increasing the expression of the OB receptor;
  - a physiologically effective amount of an OB-R agonist ligand; and a pharmaceutically acceptable excipient.
    - 29. The composition of claim 28 wherein the therapeutic cytokine capable of increasing the expression of the OB receptor and the OB-R agonist ligand are packaged separately.
- 30. The composition of claim 28 wherein the therapeutic cytokine is about 0.5 to about 20 micrograms per kilogram body weight IL-6.

- 31. The composition of claim 29 wherein the OB-R agonist ligand is administered in a dose of about 1 micrograms per kilogram body weight to about 50 micrograms per kilogram body weight recombinant human OB protein.
- 32. An assay kit for a disease marker in a sample for a systemic inflammatory response in a patient comprising:

an antibody capable of binding to OB protein; and a detection means for determining the amount of the antibody bound to OB protein.

33. A method for assaying a disease marker for an inflammatory response in a patient comprising:

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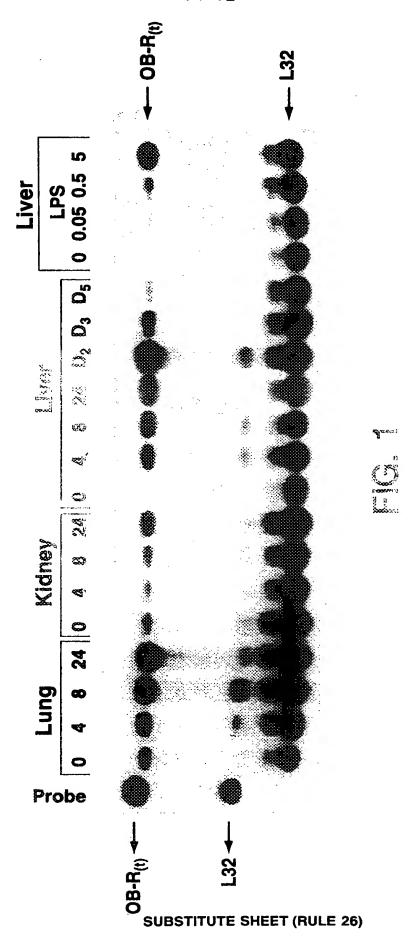
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mixing a portion of the sample with an antibody capable of binding to OB protein; and

detecting the amount of antibody bound to OB protein.

- 34. A composition suitable for the treatment of anorexia, cachexia or other wasting condition comprising a physiologically effective amount of antibody capable of binding OB protein.
- 35. The method for the treatment of anorexia, cachexia or other wasting condition comprising administering a physiologically effective amount of antibody capable of binding OB protein in an amount from about 0.02 to about 15 milligrams per kilogram body weight per day.

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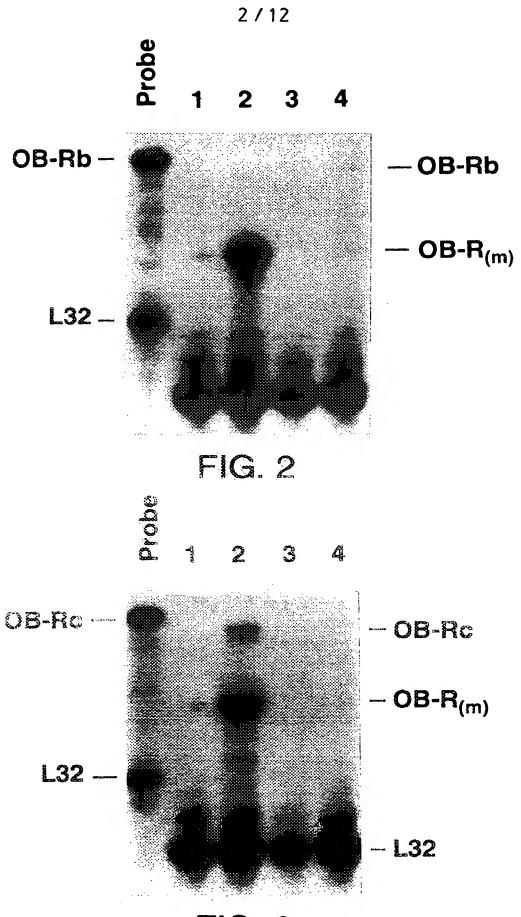
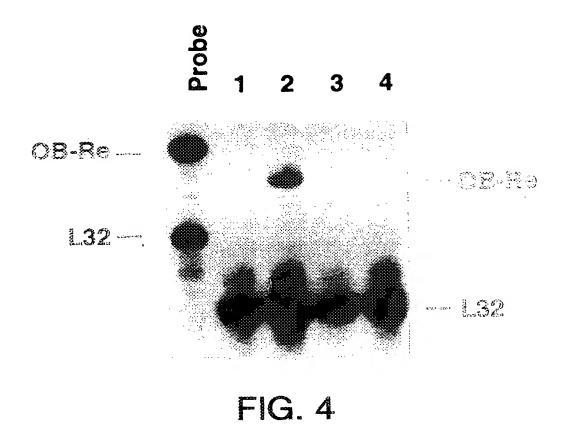


FIG. 3.
SUBSTITUTE SHEET (RULE 26)

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SUBSTITUTE SHEET (RULE 26)

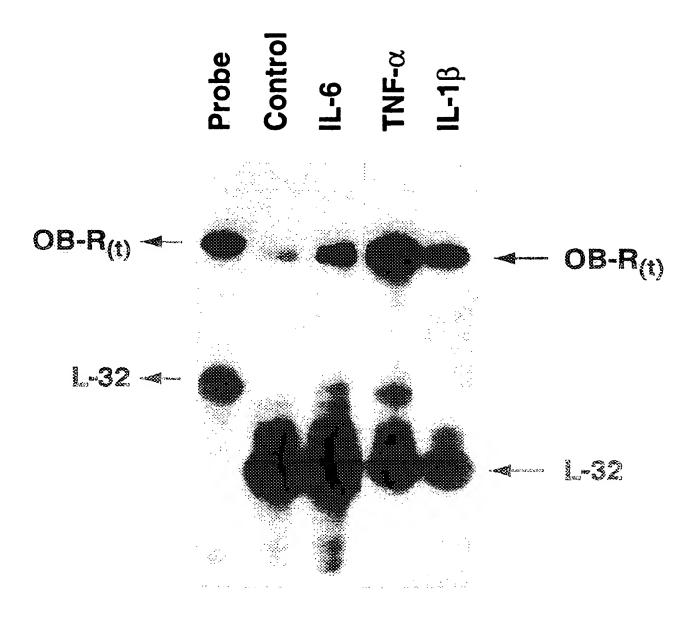
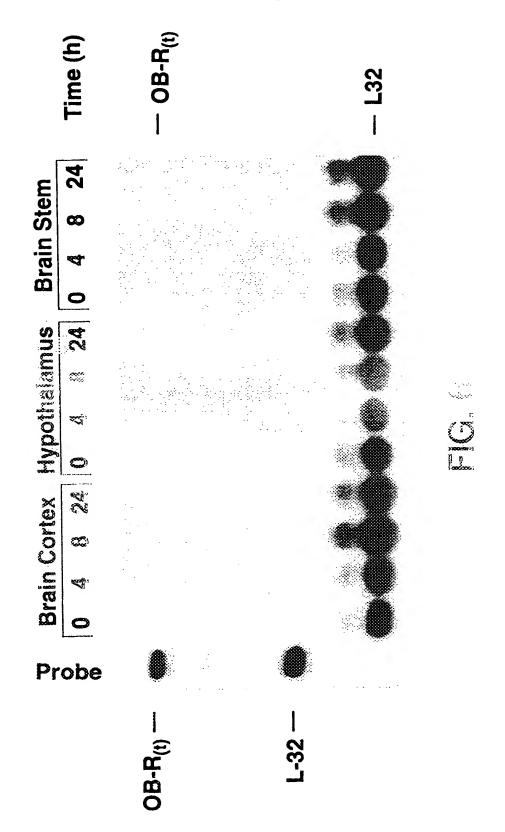


FIG. 5

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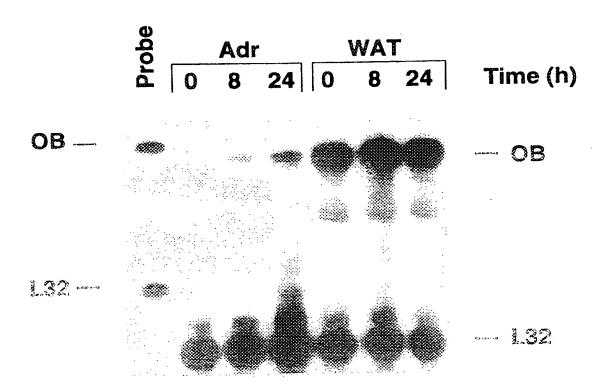


FIG. 7

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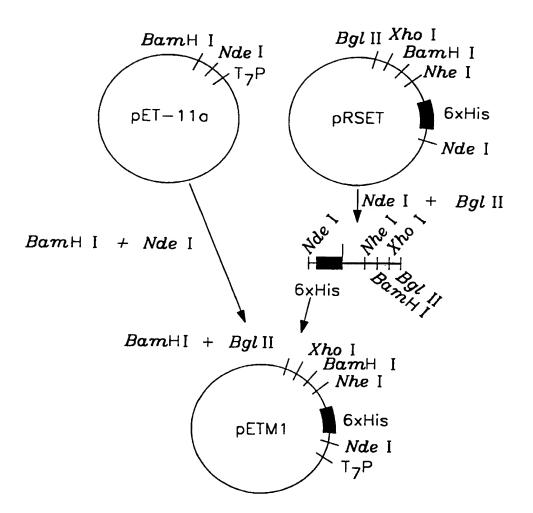


FIG. 8

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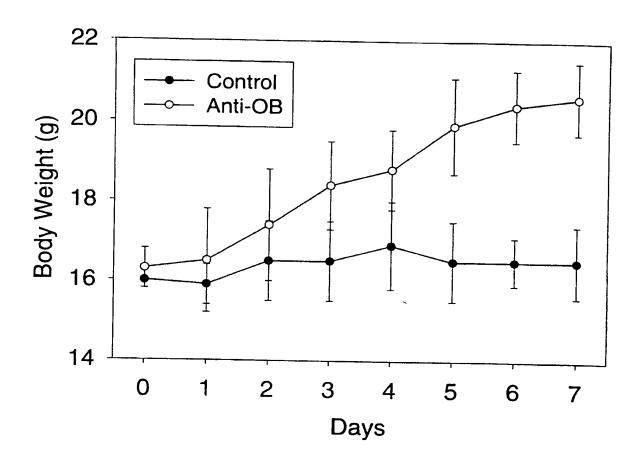


FIG. 9

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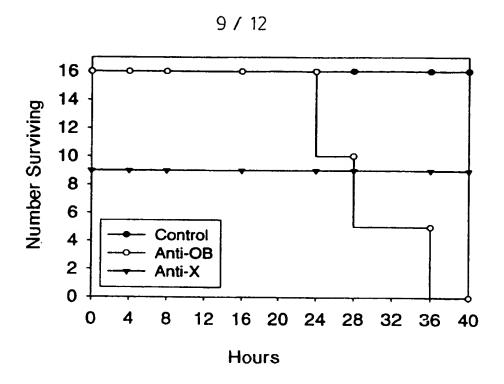


FIG. 10

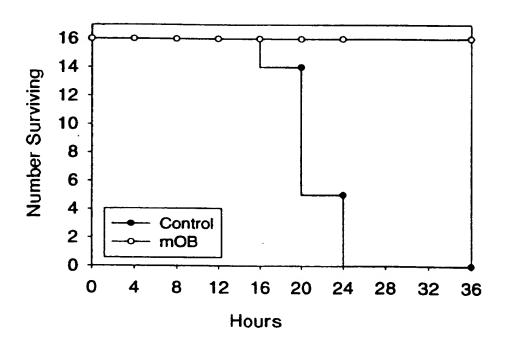


FIG. 11

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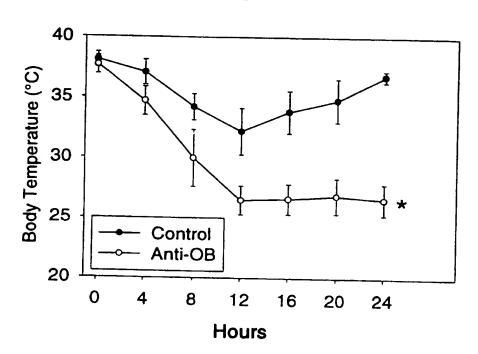


FIG. 12

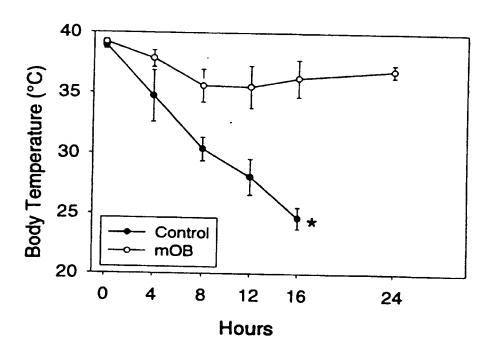


FIG. 13

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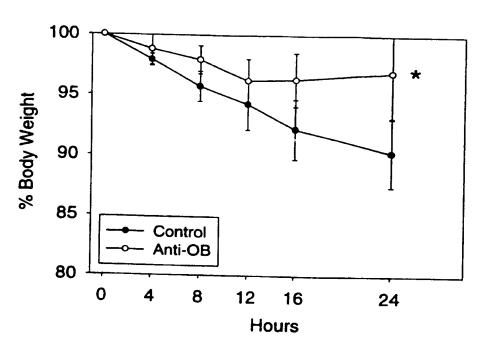


FIG. 14

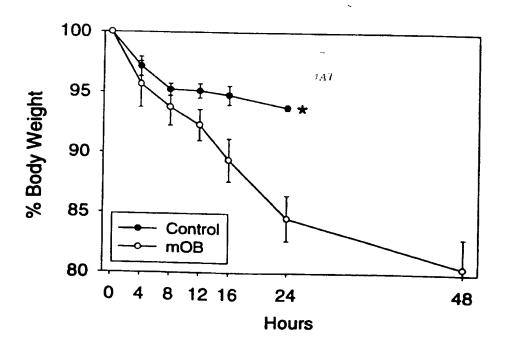
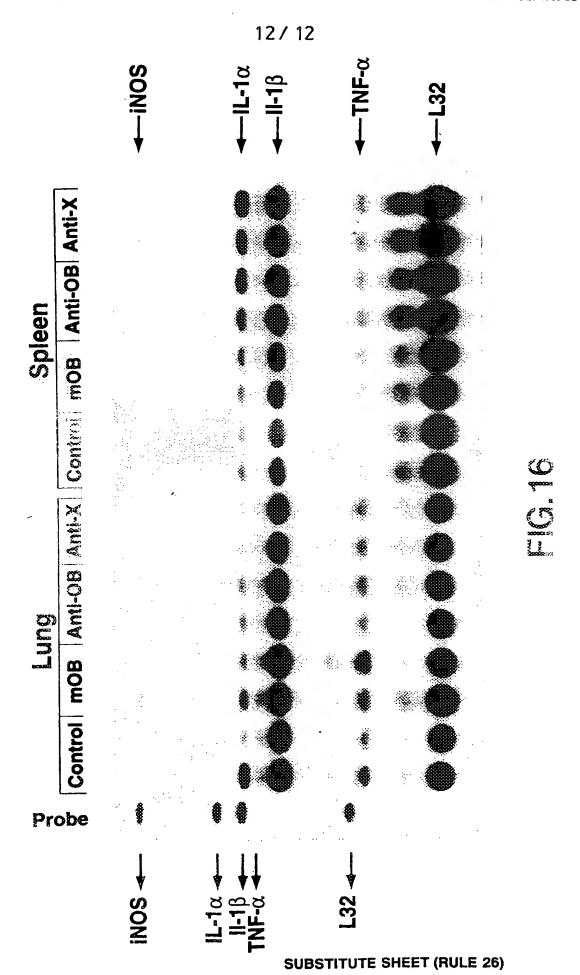


FIG. 15

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## INTERNATIONAL SEARCH REPORT

International application No.
PCT/US97/09684

			<del></del>	
A. CLASSIFICATION OF SUBJECT MATTER  IPC(6) :A61K 38/00, 45/05, 38/19, 39/395 US CL :514/2, 8, 12; 424/85.1,158.1, 145.1				
According t	to International Patent Classification (IPC) or to both	national classification	n and IPC	
	LDS SEARCHED			
Minimum d	ocumentation searched (classification system follows	d by classification sy	mbols)	
	514/2, 8, 12; 424/85.1,158.1, 145.1			
Documental	tion searched other than minimum documentation to the	e extent that such doc	uments are included	in the fields searched
Electronic d	data base consulted during the international search (r	ame of data base and	, where practicable	. search terms used)
	NLINE, DIALOG, MEDLINE		,	
	CUMENTS CONSIDERED TO BE RELEVANT			
Category*	Citation of document, with indication, where a	ppropriate, of the rele	evant passages	Relevant to claim No.
X	WO 96/05309 A3 (THE ROCKE) February 1996, (22-02-96), see t		ERSITY) 22	1-35
X, P	WO 96/35787 A1 (CHIRON CORPORATION) 14 November 1996 (14-11-96), see entire document, and claims.			1-35
X	PELLELYMOUNTER et al. Effects of the obese Gene Product on Body Weight Regulation in ob/ob Mice. Science, 28 July 1995, Vol. 269, see pages 540-543.			
X	HALAAS et al. Weight-Reducing Effects on the Plasma Protein Encoded by the obese Gene. Science, 28 July 1995, Vol. 269, see pages 543547.			8-17, 18-26
	er documents are listed in the continuation of Box (	<u> </u>	nt family annex.	
	cial categories of cited documents: cument defining the general state of the art which is not considered	date and not is	a conflict with the applica	rnational filing date or priority tion but cited to understand the
to b	be of particular relevance		ncory underlying the inve	
"L" doc	lier document published on or after the international filing date cument which may throw doubts on priority claim(s) or which is at to establish the publication date of another citation or other	considered no		claimed invention cannot be ed to involve an inventive step
spec	cial reason (as specified)	"Y" document of considered to	particular relevance; the involve an inventive	claimed invention cannot be step when the document is
"O" doc	nument referring to an oral disclosure, use, exhibition or other	combined with		documents, such combination
the	nument published prior to the internstional filing date but later than priority date claimed	*&* document men	nber of the same patent i	family
Date of the a	actual completion of the international search	Date of mailing of the	ne international sear	rch report
02 SEPTE	MBER 1997	2 3 SEP 19	<del>-</del> -	
	nailing address of the ISA/US	Authorized officer	77	
CARNETTE D. DRADED				
Facsimile No	, D.C. 20231 D. (703) 305-3230	Telephone No. (7		

## INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/09684

C (Continua	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
x	GRUNFELD et al. Endotoxin and Cytokines Induce Expression of Leptin, the ob Gene Product, in Hamsters. Journal of Clinical Investications. Clin. Invest. May 1996, Vol. 97, see pages 2152-2157.	1-35
X, P	SARRAF et al. Multiple Cytokines and Acute Inflammation Raise Mouse Leptin Levels: Potential Role in Inflammatory Anorexia. Journal of Experimental Medicine. 06 January 1997, Vol. 185, Number 1, see pages 171-175.	1-35
K, P	ROUBENOFF et al. TNF-alpha and leptin in adjuvant arthritis (AA): Implications for inflammatory cachexia. Arthritis & Rheumatism, September 1996, Vol. 39, No. 9 (supplement), see Abstract No. 302, page s77.	1-35
A, P	GAINSFORD et al. Leptin can induce proliferation, differentiation, and functional activation of hemopoietic cells. Proc. Natl. Acad. Sci. December 1996, Vol. 93, see pages 14564-14568.	1-35
A, P	BENNETT et al. A role for leptin and its cognate receptor in hematopoiesis. Current Biology, 1996, Vol. 6, No. 9, see pages 1178-1180.	1-35
	•	
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# PATENT COOPERATION TREATY

# **PCT**



# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

Applicant's or agent's file reference TSRI 540.1 PCT	FOR FURTHER ACTIO	N See Notifi Preliminary	cation of Transmittal of International Examination Report (Form PCT/IPEA/416)	
International application No.	International filing date (da	date (day/month/year) Priority date (day/month/year)		
PCT/US97/09684	04 JUNE 1997	04 JUNE 1996		
International Patent Classification (IPC) or national classification and IPC IPC(6): A61K 38/00, 45/05, 38/19, 39/395 and US Cl.: 514/2, 8, 12; 424/85.1,158.1, 145.1				
Applicant THE SCRIPPS RESEARCH INSTITUT	TE			
Examining Authority and is  2. This REPORT consists of a  This report is also accombeen amended and are th (see Rule 70.16 and Sec	total of sheets.  panied by ANNEXES, i.e., see basis for this report and/or tion 607 of the Administration	nt according to sheets of the description	ription, claims and/or drawings which have g rectifications made before this Authority.	
These annexes consist of a to	otal of sheets.			
3. This report contains indication	as relating to the followin	g items:		
I X Basis of the report  II Priority  III Non-establishment of report with regard to novelty, inventive step or industrial applicability  IV Lack of unity of invention  V X Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement  VI Certain documents cited  VII Certain defects in the international application  VIII Certain observations on the international application				
Date of submission of the demand	1	Date of completion	n of this report	
30 DECEMBER 1997		17 AUGUST	1998	
Name and mailing address of the IPEA	⁄US /	Authorized office	- A 11-	
Commissioner of Patents and Trade		GARNETTE	D DRAPER	
Washington, D.C. 20231			o far	
Facsimile No. (703) 305-3230	1	Telephone No.	(703) 308-0196 V	

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# INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No.
PCT/US97/09684

I.	Basis o	f the report		
1.	This report i	nas been drawn on the	basis of Substitute sheets	which have been furnished to the receiving Office in response to an invitation
	under Artic			iled" and are not annexed to the report since they do not contain amendments):
	x	the internations	il application as origi	nally filed.
	X	the description,	pages <u>1-38</u>	, as originally filed.
				, filed with the demand.
				, filed with the letter of
			pages	, filed with the letter of
	Ιx	the claims,	Nos. <u>1-35</u>	, as originally filed.
	_	•	Nos. NONE	, as amended under Article 19.
				, filed with the demand.
				, filed with the letter of
			Nos	, filed with the letter of
	x	the drawings,	sheets/fig 1-16	, as originally filed.
				, filed with the demand.
				, filed with the letter of
			sheets <del>/fig</del>	, filed with the letter of
	x x	the claims,	Nos. NONE  Nos. NONE  sheets/fig NONE	
3	· T	his report has been on go beyond the disc	established as if (some of osure as filed, as indicated)	of) the amendments had not been made, since they have been considered ted in the Supplemental Box Additional observations below (Rule 70.2(c)).
4	I. Additio	nal observations,	if necessary:	
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### INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No.

PCT/US97/09684

V.	R asoned stat ment und r Articl	35(2) with regard t	novelty, inventive step or	industrial applicability;
	citati ns and explanati ns support	ing such stat ment		

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#### 2. CITATIONS AND EXPLANATIONS

Claims 8-17 lack novelty under PCT Article 33(2) as being anticipated by, or in the alternative lack an inventive step under PCT Article 33(3) as being obvious over The Rockefeller University ('309), Grunfeld et al, Pelleymounter et al or Halaas et al.

The claims are directed to compositions of the Ob protein or an Ob agonist ligand, wherein the claims recited several intended uses for the composition such as regulating energy metabolism during a systemic inflammatory response, or for ameliorating the toxicity of cytokines. Each of the prior art disclose composition of the Ob protein or related protein that are agonist, but the protein is disclosed for use in a different method from those of the claims. The prior art anticipate or render obvious the claims for a composition of the Ob protein irrespective of statements of intended for the composition. Furthermore, a composition per se is not distinct even if it is

used for a different purpose or in a different method. Therefore, the burden is on the applicants to prove that the prior art composition does not possess the properties for methods of treatment consistent with the claims.

Claims 32-35 lack novelty under PCT Article 33(2) as being anticipated by, or in the alternative lack an inventive step under PCT Article 33(3) as being obvious The Rockefeller University ('309).

In addition to the above teachings, Friedman et al also disclose antibodies to the ob protein and teach that these antibodies can be used in diagnostic methods to detect for disorders associated with the Ob protein. Thus, the prior art appears to meet the claim limitation, irrespective of the fact that the language used to define the invention differs, and irrespective of statements of intended use for the antibody composition. Also taught is that the antibodies can be used to antagonist the undesired effect of the leptin, therefore to use the antibodies to treat low weight disorders is also anticipated or obvious from such teachings.

Claims 28-31 lack novelty under PCT Article 33(2) as being (Continued on Supplemental Sheet.)

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### INTERNATIONAL PRELIMINARY EXAMINATION REPORT

International application No.

PCT/US97/09684

Supplem n	tal Bo	Z
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(To be used when the spac in any of the preceding boxes is not sufficient)

Continuation of: Boxes I - VIII

Sheet 10

#### V. 2. REASONED STATEMENTS - CITATIONS AND EXPLANATIONS (Continued):

anticipated by, or in the alternative lack an inventive step under PCT Article 33(3) as being obvious Grunfeld et al.

Similar to the above, these claims are directed to a composition of the Ob protein and a cytokine, and wherein the claims recite specific statements of intended use for the composition. Grunfeld et al teach that endotoxin and cytokines induce the expression of leptin/ob in response to infection, and with this discovery, they provide compositions of the Ob protein and cytokines such as TNF, IL-1, Il-6, Il-8 and IFN. The prior art anticipate or render obvious the claims for a composition of the Ob protein and a cytokine irrespective of statements of intended for the composition. Purthermore, the composition per se is not distinct even if it is used for a different purpose or in a different method. Therefore, the burden is on the applicants to prove that the prior art composition does not possess the properties for methods of treatment consistent with the claims.

Claims 1-7, 18-31 lack an inventive step under PCT Article 33(2) as being obvious Grunfeld et al.

The disclosure of Grunfeld et al has been set forth above. Since the prior art teach that endotoxin such as LPS and cytokines induce expression of the Ob protein in response to infection. In view of the well known fact that many infections are associated with or are accompanied by inflammations, at the time of the invention it would have been obvious to use the ob protein alone or in conjunction with the endotoxin or a cytokine to regulate energy metabolism during inflammation that are associated with the ob protein.

Claims 1-35 the criteria set out in PCT Article 33(4); and claims 1-7 and 18-27 meet the criteria set out in PCT Article 33(2) because the prior art does not expressly teach the use of an Ob-R inducer with a cytokine and the Ob/leptin to treat obesity as claimed.

	NEW CITATIONS	
NONE		

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# **PATENT COOPERATION TREATY**

From t	he IN	TERNA'	TIONAL	BUREAU
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PCT	To:
NOTIFICATION OF ELECTION  (PCT Rule 61.2)	United States Patent and Trademark Office (Box PCT) Crystal Plaza 2 Washington, DC 20231 ETATS-UNIS D'AMERIQUE
Date of mailing (day/month/year) 16 February 1998 (16.02.98)	in its capacity as elected Office
International application No. PCT/US97/09684	Applicant's or agent's file reference TSRI 540.1 PCT
International filing date (day/month/year) 04 June 1997 (04.06.97)	Priority date (day/month/year) 04 June 1996 (04.06.96)
Applicant FENG, Lili et al	
1. The designated Office is hereby notified of its election made  X in the demand filed with the International Preliminary  30 December  in a notice effecting later election filed with the International Preliminary  The election X was  was not  made before the expiration of 19 months from the priority of Rule 32.2(b).	r Examining Authority on:  1997 (30.12.97)  national Bureau on:
The International Bureau of WIPO 34, chemin des Colombettes 1211 Geneva 20, Switzerland	Authorized officer  J. Leitao

Telephone No.: (41-22) 338.83.38

Facsimile No.: (41-22) 740.14.35

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## yon for the Elected Office (EO/US)

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### From the INTERNATIONAL BUREAU PCT To: NOTIFICATION OF THE RECORDING CEPURITIS, Talivaldis **OF A CHANGE** Olson & Hierl, Ltd. 36th floor (PCT Rule 92bis.1 and 20 North Wacker Drive Administrative Instructions, Section 422) Chicago, IL 60606 **ETATS-UNIS D'AMERIQUE** Date of mailing (day/month/year) 20 February 1998 (20.02.98) Applicant's or agent's file reference IMPORTANT NOTIFICATION **TSRI 540.1 PCT** International filing date (day/month/year) International application No. 04 June 1997 (04.06.97) PCT/US97/09684 1. The following indications appeared on record concerning: the common representative the agent the inventor the applicant State of Residence State of Nationality Name and Address US CN CHEN, Sizhong 7896 Camino Kiosco San Diego, CA 92122 Telephone No. United States of America Facsimile No. Teleprinter No. 2. The International Bureau hereby notifies the applicant that the following change has been recorded concerning: the residence the nationality the address the person X the name State of Residence State of Nationality Name and Address US CN CHEN, Shizhong Telephone No. Facsimile No. Teleprinter No. 3. Further observations, if necessary: 4. A copy of this notification has been sent to: the designated Offices concerned the receiving Office the elected Offices concerned the International Searching Authority other: the International Preliminary Examining Authority Authorized officer The International Bureau of WIPO 34, chemin des Colombettes Aino Metcalfe 1211 Geneva 20, Switzerland Telephone No.: (41-22) 338.83.38 Facsimile No.: (41-22) 740.14.35

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### INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/09684

-	SSIFICATION OF SUBJECT MATTER A61K 38/00, 45/05, 38/19, 39/395					
US CL :	514/2, 8, 12; 424/85.1,158.1, 145.1					
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Documentati	ion searched other than minimum documentation to th	e extent tha	t such docun	nents are included	in the fields searched	
	ata base consulted during the international search (n LINE, DIALOG, MEDLINE	ame of data	a base and, v	where practicable,	, search terms used)	
C. DOC	UMENTS CONSIDERED TO BE RELEVANT					
Category*	Citation of document, with indication, where a	ppropriate,	of the releva	ant passages	Relevant to claim No.	
X	WO 96/05309 A3 (THE ROCKERFELLER UNIVERSITY) 22 1-35 February 1996, (22-02-96), see the claims.					
X, P	WO 96/35787 A1 (CHIRON CORPORATION) 14 November 1-35 1996 (14-11-96), see entire document, and claims.					
x	PELLELYMOUNTER et al. Effects of the obese Gene Product on Body Weight Regulation in ob/ob Mice. Science, 28 July 1995, Vol. 269, see pages 540-543.					
×	HALAAS et al. Weight-Reducin Protein Encoded by the obese Gen Vol. 269, see pages 543547.		8-17, 18-26			
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X Furth	er documents are listed in the continuation of Box C	z. 🔲	See patent	family annex.		
"A" doc	cial categories of cited documents:  ument defining the general state of the art which is not considered se of particular relevance		date and not in o	published after the inteconflict with the applications underlying the investment	rnational filing date or priority tion but cited to understand the ention	
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Washington Facsimile No	, D.C. 20231 p. (703) 305-3230	GARNETTE D. DRAPER Telephone No. (703) 308-0196				
		relephone	, 140. (70	3) 300-0190		





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International application No. PCT/US97/09684

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
x	GRUNFELD et al. Endotoxin and Cytokines Induce Expression of Leptin, the ob Gene Product, in Hamsters. Journal of Clinical Investications. Clin. Invest. May 1996, Vol. 97, see pages 2152-2157.	1-35
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А, Р	BENNETT et al. A role for leptin and its cognate receptor in hematopoiesis. Current Biology, 1996, Vol. 6, No. 9, see pages 1178-1180.	1-35
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